The development of vaccine candidates against Plasmodium vivax—the most geographically widespread human malaria species—is challenged by technical difficulties, such as the lack of in vitro culture systems and availability of animal models. Chimeric rodent Plasmodium parasites are safe and useful tools for the preclinical evaluation of new vaccine formulations. We report the successful development and characterization of chimeric Plasmodium berghei parasites bearing the type I repeat region of P. vivax circumsporozoite protein (CSP). The P. berghei-P. vivax chimeric strain develops normally in mosquitoes and produces highly infectious sporozoites that produce patent infection in mice that are exposed to the bites of as few as 3 P. berghei-P. vivax-infected mosquitoes. Using this transgenic parasite, we demonstrate that monoclonal and polyclonal antibodies against P. vivax CSP strongly inhibit parasite infection and thus support the notion that these antibodies play an important role in protective immunity. The chimeric parasites we developed represent a robust model for evaluating protective immune responses against P. vivax vaccines based on CSP.

The circumsporozoite protein (CSP) is the most abundant protein on the surfaces of sporozoites and is responsible for eliciting both T-cell and antibody responses. The plasmoidal CSP is one of the best studied antigens and is considered to be a major malaria vaccine candidate. RTS,S, the most effective malaria vaccine candidate to date, is based on P. falciparum CSP, thus underscoring CSP’s immunogenic properties. CSP is comprised of an immunodominant central repeat region, diverse among Plasmodium species, flanked by two conserved domains: region I at the N terminus of the repeats and a type I thrombospondin repeat (TSR) motif C terminal to the repeat region. Early studies in animals have indicated that antibodies against the repeat region neutralize the infectivity of sporozoites and confer sterile immunity (11, 12). Importantly, recent studies in humans vaccinated with RTS,S indicated that protection among vaccinees correlates with the titer of anti-CSP repeat antibodies (13).

Plasmodium berghei parasites bearing the repeat region of P. falciparum CSP have been developed and characterized by Persson et al. (14). These parasites, “antigenically P. falciparum but functionally rodent malaria” (14), are a practical tool for evaluating the efficacy of human preerythrocytic P. falciparum CSP-based vaccine candidates. On the basis of the study by Persson et al., we developed and characterized a chimeric P. berghei parasite strain expressing the repeat region of P. vivax (VK210) CSP as a first step in evaluating vaccine candidates based on the CSP of P. vivax. We demonstrate that these chimeric parasites are recognized by a monoclonal antibody (MAb), 2F2, specific for the P. vivax VK210 CSP repeats (15) and that the in vivo infectivity of chimeric sporozoites is strongly inhibited by this monoclonal antibody, as well as by polyclonal antibodies raised against a recombinant P. vivax CSP construct (16).

MATERIALS AND METHODS

Plasmid construction and parasite transfection. The transfection plasmid, pR-CSPv, carrying the repeat region of P. vivax, was derived from plasmid p8-CSwt (17), which carries the CS gene of P. berghei (wild type [WT]) and results from the combination of plasmids pIC-CSwt (18) and pPv (Retrogen Inc., CA) encoding the VK210 repeat motif based on the Salvador I (Sall) strain of P. vivax. Briefly, a PmlI-SexAI 567-bp fragment was excised from pIC-CSwt and replaced with a PmlI-SexAI 675-bp frag-
ment comprising the P. vivax VK210 CSP repeat region, which was codon harmonized to optimize protein synthesis and folding in the P. berghei transgenic host (19) and was excised from the engineered pPV. From the resulting intermediate plasmid, p-CSVp, the entire CSV gene was excised as a KpnI-XhoI fragment and inserted into the transfection plasmid, p-R-CSVp. KpnI and SacI were used to linearize p-R-CSVp prior to transfection; WT P. berghei (strain ANKA) parasites were transfected as previously described (20).

Chimeric parasites were selected in Swiss Webster mice (NCI, Frederick, MD) by treatment with pyrimethamine in drinking water (0.07 mg/ml). Parasites surviving drug treatment were then cloned by means of a limiting-dilution assay (21). Recombination at the 3’ and 5’ ends of the locus was verified by PCR. The primers used to verify integration at the 5’ end of the construct are 5’F (TCACCCTCAAGTTGGGTAAAA) and 5’R (CCTGTCCCTGTTGTGCTA); the primers to verify integration at the 3’ end are 3’F (TGTAATAATGTGTAGTGTGC) and 3’R (GTGCCCAATTCGACTTGC). To verify that the isolated clone did not have contaminating WT P. berghei parasites, a PCR product flanking the SexAI restriction site was digested with SexAI. The primers used for this PCR are PBs-F (AAGAACGAAGATTGAGCCT) and PBs-C-R (AGAAGTCAAAACTCTGCTTCTTCTT). Finally, DNA isolated from the cloned transgenic parasites was sequenced (Macrogen Inc., USA) to confirm the replacement of the P. berghei repeat region with the P. vivax repeat region for the generation of P. berghei-P. vivax chimeric parasites.

Mosquito infection and parasite development. Anopheles stephensi mosquitoes were fed on Swiss Webster mice (NCI, Frederick, MD) infected with blood stages of cloned P. berghei-P. vivax CSRepeat (P. berghei-P. vivax) parasites. Development of viable male gametocytes in blood was qualitatively assessed by means of an exflagellation assay. Briefly, 1.5 μl of blood from a mouse with patent blood parasitemia was incubated at room temperature for 20 min with 1.5 μl of heparin (40 U/ml in 1× phosphate-buffered saline [PBS]) and 7 μl of complete ookinet medium (RPMI 1640 medium with 10% fetal calf serum, 25 mM HEPES, 2 mM glutamine, 0.2 mM pyruvate, 12 mM sodium bicarbonate, 1 μM xantheneuric acid). After incubation, the mixture was placed on a slide and observed under a light microscope (×100 magnification).

Development of oocysts in mosquito midguts was assessed 14 days post-blood meal. The midguts were dissected in sterile PBS and stained in a 0.1% mercury chromate solution.

Infectivity of P. berghei-P. vivax sporozoites. The in vivo infectivity of chimeric P. berghei-P. vivax sporozoites was assessed in 5- to 8-week-old C57BL/6 mice both by assessing the liver parasite load, measured by quantitative PCR (qPCR) (22), in mice that were challenged intravenously (i.v.) with 1 × 10^7 P. berghei-P. vivax sporozoites and by the development of blood-stage parasites after feeding with 3 P. berghei-P. vivax infected A. stephensi mosquitoes. For increased stringency, we allowed 4 mosquitoes to feed for 5 min and then visually examined them for a blood feed. The presence of sporozoites was then assessed in those mosquitoes that fed to ensure that mice were exposed to the bite of at least 1 infected mosquito.

Data analysis. Most of the data were plotted using Prism 4 software (GraphPad). Unless otherwise stated, data were compared for significance using a Mann-Whitney test.

RESULTS

Generation and biological characterization of P. berghei-P. vivax chimeric parasites. Using a plasmid containing the sequences of the P. berghei CSP gene and a modified version of the dihydrofolate reductase (DHFR) gene, which confers resistance to pyrimethamine on transfected parasites, we generated P. berghei-P. vivax chimeric parasites. Briefly, we replaced the repeat region of P. berghei CSP with the repeat domain of P. vivax CSP. The P. vivax replacement represents the VK210 CSP sequence of the P. vivax SalI strain, from nucleotides 268 to 825 (amino acids 90 to 275). P. berghei ANKA blood stages were transfected in vitro with a plasmid linearized using KpnI and XhoI restriction enzymes, as described previously (21) (Fig. 1A). The transfected parasites were injected intravenously into Swiss Webster mice, and transgenic parasites were selected under pyrimethamine treatment. Ten days later, drug-resistant parasites were recovered and cloned by limiting dilution.

PCR analysis was used to confirm recombination at the 5’ and 3’ ends for gene replacement (Fig. 1B), and the cloned parasites were verified by PCR-restriction fragment length polymorphism (RFLP) (Fig. 1C). The cloned parasites were sequenced to further confirm that the P. berghei-P. vivax parasites have the correct insertion in the CSP gene (Fig. 1D).

Mosquito infection. A cloned P. berghei-P. vivax parasite isolate was used to infect mice, which were then used to feed A. stephensi mosquitoes. Transgenic parasites were evaluated by using standard methods, such as exflagellation and oocyst and sporozoite counts, to compare them to the wild-type P. berghei.

Exflagellation of male P. berghei-P. vivax gametocytes, assessed prior to feeding female A. stephensi mosquitoes on infected mice, was normal and comparable to that of WT P. berghei ANKA parasites (data not shown). Twelve to 14 days after mosquito feeding, P. berghei-P. vivax-infected mosquito midguts had approximately 100 to 150 healthy-looking oocysts per midgut, which was also comparable to WT P. berghei ANKA parasites. Finally, on day 21, mosquito salivary glands were dissected, and the parasite load was found to be comparable to that of WT P. berghei ANKA (Table 1).
Characterization of *P. berghei*-*P. vivax* sporozoites with monoclonal antibodies. Sporozoites obtained from salivary glands of *P. berghei*-*P. vivax*-infected mosquitoes were used for IFA with antibodies specific for the repeat domain of *P. vivax* (2F2) CSP. An antibody specific for the repeat region of *P. berghei* (3D11) served as a control. The IFA results clearly demonstrated that the chimeric *P. berghei*-*P. vivax* sporozoites were all uniformly positive for the anti-*P. vivax* repeat antibodies and negative for the antibody against *P. berghei* repeats (Fig. 2).

Infectivity of *P. berghei*-*P. vivax* chimeric parasites in mice. To evaluate the *in vivo* fitness of the transgenic parasites, we challenged C57BL/6 mice with $1 \times 10^4$ sporozoites, delivered by i.v. injection, from either WT *P. berghei* ANKA or *P. berghei*-*P. vivax* parasites. Forty-two hours after infection, the parasite load in the liver was measured by qPCR specific for the *P. berghei* 18S rRNA.

**TABLE 1** Developmental characteristics and infectivity of *P. berghei*-*P. vivax* parasites in *A. stephensi* mosquitoes

| Parasite | % of midgut infected | No. of oocysts/midgut | % of salivary gland infected | No. of sporozoites/salivary gland ($10^3$) | % of mice infected after mosquito bites$^b$
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<tr>
<td><em>P. berghei</em>-<em>P. vivax</em></td>
<td>80.72</td>
<td>54.76</td>
<td>75</td>
<td>8.5</td>
<td>100</td>
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<tr>
<td><em>P. berghei</em> ANKA</td>
<td>75.14</td>
<td>42.25</td>
<td>66</td>
<td>10</td>
<td>100</td>
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$^a$ Mean of 3 experiments, with at least 20 mosquitoes examined in each experiment.

$^b$ Three infected mosquitoes were allowed to feed on C57BL/6 mice for 3 min. Blood-stage parasitemia was assessed 5 days later.
The data indicated that the infectivities of WT *P. berghei* ANKA and *P. berghei*-*P. vivax* sporozoites were comparable, and there was no statistically significant difference between the parasites. The infectivity of the transgenic parasites was further confirmed by delivering the parasites via the natural route, i.e., using mosquito bites. Mice exposed to the bites of 3 infected mosquitoes demonstrated patent blood-stage infection by day 5 postchallenge.

**Anti-*P. vivax* antibodies inhibit the development of parasite liver stages.** In order to evaluate the utility of the transgenic parasites as a tool to assess the efficacy of future vaccines, we tested the neutralization capabilities of anti-*P. vivax* antibodies *in vivo*. The anti-repeat monoclonal antibody 2F2 was passively transferred into mice immediately prior to an intravenous challenge with $1 \times 10^4$ chimeric *P. berghei*-*P. vivax* sporozoites. As seen in Fig. 3, mice that received 2F2 antibody showed a significant reduction in parasite load in the liver (99% reduction compared to naive controls). In contrast, the group that received the irrelevant antibody had a parasite load comparable to that of naive controls, confirming the specificity of the results observed with the anti-*P. vivax* MAb 2F2.

To determine the effects of polyclonal antibodies that contain a mixture of specificities (as opposed to the repeat-specific monoclonal antibody 2F2), we evaluated the neutralization capability of rabbit polyclonal serum that was generated against a recombinant *P. vivax* CSP protein, VMP001. Rabbit anti-VMP001 antibodies recognized CSP on the surfaces of *P. vivax* sporozoites and reacted to the whole protein (VMP001), in addition to the N-terminal and C-terminal proteins, as well as a synthetic type 1 peptide. As shown in Fig. 3B, similar to MAb 2F2, polyclonal rabbit sera also reduced the liver parasite burden in comparison to mice that either were naive or received preimmune rabbit serum. When mice receiving 400 µg of monoclonal antibody 2F2 were exposed to the bites of 4 *P. berghei*-*P. vivax*-infected mosquitoes for 5 min, we did not observe sterile protection. However, control mice ($n = 5$) receiving an irrelevant monoclonal antibody developed parasitemia at day 4, while mice receiving anti-parasite monoclonal antibody ($n = 5$) became patent at day 5. Taken together, these results demonstrate that *P. berghei*-*P. vivax* sporozoites are highly infectious and a stringent model to evaluate immune responses.
that inhibit parasite development in the liver, as well as the acquisition of sterile immunity after immunization with vaccine candidates.

**DISCUSSION**

Studies on assessing protective immune responses against malaria in general, and *P. vivax* in particular, are difficult due to the inability to culture the parasite and to the lack of a suitable animal model. Human studies are expensive, and therefore, this is not a practical first step to evaluate vaccine candidates. Over the last decade or so, transfection of *Plasmodium* has revolutionized the ways in which protective immune responses can be evaluated using transgenic rodent parasites that have been transfected with the gene encoding a specific human malaria antigen (14, 26–29). Active- and passive-immunization studies directed toward these antigens are more indicative of a biologically relevant immune response than the evaluation of immunogenicity alone. Transgenic parasites carrying the CSP, MSP, and P25 genes of *P. falciparum* have been developed and are being used to evaluate vaccine candidates and/or platforms (14, 26–29). To date, only one transgenic *P. berghei* sporozoite expressing P25 from *P. vivax* has been developed as a tool for evaluating the efficacy of vaccine candidates (29).

Here, we describe the successful development, characterization, and applicability of chimeric *P. berghei* parasites expressing the repeat region of *P. vivax* CSP. We successfully replaced the *P. berghei* repeat region with the VK210 repeat region of *P. vivax*. The resulting parasites produce viable gametocytes and successfully infect *A. stephensi* mosquitoes, in which their development is comparable to that of WT *P. berghei*. The *in vivo* infectivity of *P. berghei-P. vivax* sporozoites is comparable to that of WT *P. berghei* in C57BL/6 mice. Moreover, feeding of *P. berghei-P. vivax* infected mosquitoes on naive C57BL/6 mice results in the development of patent blood-stage parasitemia. Following immunization, the true potential of a vaccine needs to be evaluated using a natural route of sporozoite inoculation, which allows the participation of all components of the immune response in protection.

We used the transgenic parasites to evaluate their applicability as a tool for assessing the efficacies of antibodies in protection. Passive transfer of monoclonal antibodies or hyperimmune sera into naive recipients conferred significant protection against *P. berghei-P. vivax* sporozoite challenge. Mice that received 2F2 antibody or hyperimmune rabbit sera both showed above 95% inhibition of the parasite burden in the liver compared to the naive controls, thus proving that the transgenic parasites can serve as an important tool in evaluating protective anti-repeat immune responses. Given that sporozoites rapidly migrate through the circulatory system (30), the availability and affinity of circulating anti-sporozoite antibodies determine their protective capacity. While the monoclonal antibody 2F2 was able to decrease liver infection by 90% of the sporozoites—as shown by measurement of the parasite load in the liver—and delayed the prepatent period by 1 day, it does not appear to be efficient enough to confer sterile immunity. Further comparative studies will be necessary to define the dosage, avidity, and fine specificity of different monoclonal and polyclonal antibodies capable of conferring sterile immunity. The possibility that sterile immunity was not achieved due to generation of mutants or revertant parasites is unlikely; however, this is an issue worth considering for future studies. Taken together, these results demonstrate that the *P. berghei-P. vivax* parasites are robust and that a natural mosquito bite challenge can be used as a stringent test to assess the full potential of vaccine candidates. Passive-transfer experiments in mice using serum samples from volunteers immunized with novel vaccine candidates could potentially be used to assess humoral protective capacity against *P. berghei-P. vivax* sporozoites. Such studies should also be useful for determining more accurate correlates between antibody titers and protective efficacy against *P. vivax* infection.

Chimeric rodent malaria parasites are a simple, robust, and practical tool for the in vivo evaluation of vaccine candidates against malaria. This is the first report of the development of a rodent parasite that can be used to evaluate the efficacy of the circumsporozoite protein central repeat region of *P. vivax*. As the replacement of the entire CSP gene could result in parasites with reduced infectivity in both mice and mosquitoes (31), we chose to first develop chimeric parasites in which we replaced only the *P. berghei* CSP repeat domain with that of *P. vivax*, which is a well-defined target of protective antibody responses. The successful generation of parasites that are able to reproducibly infect mice opens up the road to develop additional transgenic parasites expressing the N-terminal and C-terminal regions and, if possible, the full-length CSP to evaluate T-cell-mediated immune-protective mechanisms.

Such parasites will help select future vaccine candidates for clinical development. Over the last decade, the utility of chimeric rodent parasites has been highlighted by an increasing number of studies that report their use for the assessment of novel protein constructs, recombinant viral vectors, and synthetic adjuvant formulations (32–37). The *P. berghei-P. vivax* parasites we have generated should also facilitate the appraisal of new vaccine candidates and represent a practical platform for evaluating CSP-based protective immune responses against *P. vivax*.

**ACKNOWLEDGMENTS**

F.Z. is supported by National Institutes of Health grant AI44375. We are grateful for the support of the Bloomberg Family Foundation.

The opinions or assertions contained herein are our private views and are not to be construed as official or as reflecting the views of the Department of the Army or the Department of Defense.

**REFERENCES**


