Contribution of *Salmonella typhimurium* Virulence Factors to Diarrheal Disease in Calves

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Limited knowledge is available about the virulence mechanisms responsible for diarrheal disease caused by *Salmonella typhimurium*. To assess the contribution to diarrheal disease of virulence determinants identified in models of infection, we tested a collection of *S. typhimurium* mutants for their ability to cause enteritis in calves. *S. typhimurium* strains carrying mutations in the virulence plasmid (*spv*), *Salmonella* pathogenicity island 2 (*SPI-2*) (*spIB*), or *SPI-5* (*sopB*) caused mortality and acute diarrhea in calves. An *S. typhimurium* *rfaJ* mutant, which is defective for lipopolysaccharide outer core biosynthesis, was of intermediate virulence. Mutations in *SPI-1* (*hilA* and *prgH*) or *aroA* markedly reduced virulence and the severity of diarrhea. Furthermore, histopathological examination of calves infected with *SPI-1* or *aroA* mutants revealed a marked reduction or absence of intestinal lesions. These data suggest that virulence factors, such as *SPI-1*, which are required during intestinal colonization are more important for pathogenicity in calves than are genes required during the systemic phase of *S. typhimurium* infection, including *SPI-2* or the *spv* operon. This is in contrast to the degree of attenuation caused by these mutations in the mouse.

*Salmonella typhimurium* has a murine reservoir (10) where it produces a typhoid fever-like disease (33). After oral infection of mice, bacteria invade the mucosa of the small intestine, displaying a tropism for M cells located in the follicle-associated epithelium of Peyer’s patches (21). Upon intestinal penetration, *S. typhimurium* causes a transient bacteremia and seeds systemic sites of infection, most importantly the liver and the spleen. *S. typhimurium* multiplies in splenic and hepatic tissues at a net growth rate of 0.5 to 1.5 log/day (19, 28). After bacterial numbers in liver and spleen reach between 10⁷ and 10⁹ CFU/organ, mice die from lipopolysaccharide (LPS)-induced damage of liver and spleen, which appears to be distinct from endotoxic shock (24).

In addition to causing illness in mice, *S. typhimurium* is frequently associated with disease in humans and livestock (42, 43, 45). The illness caused by *S. typhimurium* in these host species differs markedly, however, from murine typhoid. First, mice do not develop diarrhea, the prominent sign of disease during *S. typhimurium* infections in livestock and humans. Second, *S. typhimurium* infections in humans and livestock usually remain localized to the small intestine and mesenteric lymph node, while mice develop a systemic infection. The differences between murine typhoid and diarrheal disease raise the question whether *S. typhimurium* virulence mechanisms identified in the mouse are of equal importance during infections of humans or livestock.

*S. typhimurium* is a major cause of calf morbidity and mortality in the United States and in Europe (36, 40, 44). Furthermore, the infection in calves closely resembles illness caused by *S. typhimurium* in humans and can therefore serve as a model to study diarrheal disease. Calves infected experimentally with *S. typhimurium* develop diarrhea within 48 h (34). The bacteria invade the intestinal epithelium in the terminal ileum, resulting in exfoliation of epithelial cells and stunting of villi (11). Bovine enteritis caused by *S. typhimurium* is primarily an enteric infection with mortality resulting from dehydration and intestinal lesions. At necropsy of terminally infected calves, acute fibrinopurulent necrotizing enteritis is visible in both the villous ileum and Peyer’s patches (53). Histopathological examination reveals destruction of the mucosal epithelium, massive neutrophil infiltration, and depletion of lymphocytes in the germinal centers of intestinal lymphoid follicles (53).

At least 60 genes are required for full virulence of *S. typhi-
murium* in mice (15), but the roles of a only few of them, namely, *aroA*, *dap*, *galE*, and *invH*, during oral infection in cattle have been studied. Following oral inoculation of calves, *S. typhimurium* *invH*, *dap*, or *aroA* mutants show a marked reduction in the severity of enteritis (7, 46, 51). *S. typhimurium* *aroA* or *aroA* *aroD* mutants are furthermore attenuated sufficiently to be used as oral vaccines in calves (23, 46). Mutational inactivation of *galE*, on the other hand, results in attenuation of *S. typhimurium* in calves but does not render this pathogen unable to cause mortality when administered orally at a high dose (6). *S. typhimurium* strains carrying mutations in *aroA*, *galE*, *dap*, or *invH* elicit reduced fluid accumulation in bovine ligated ileal loops compared to that elicited by the wild type, suggesting that this assay is predictive of attenuation in calves (8, 51). Other mutations which reduce the ability of *S. typhimurium* to elicit fluid secretion in bovine intestinal loops include those in *sirA* and *hilA* (1). Furthermore, mutations in *prgH*, *hilA*, *rfal*, *sopR*, or *spIB* reduce the ability of *S. typhimurium* to colonize bovine tissues during competitive infection experiments (49). However, it is not clear from these experiments to what degree mutations in *sirA*, *prgH*, *hilA*, *rfal*, *sopR*, or *spIB* reduce *S. typhimurium* virulence or the severity of intestinal lesions during an oral infection of calves.

A second *Salmonella* serotype frequently associated with disease in cattle is *Salmonella dublin* (14, 44). Several loci, including *aroA*, *invH*, and the *spv* operon, contribute to virulence of *S. dublin* during oral infection of calves (26, 30, 37, 50,
Furthermore, mutations in the *S. dublin invH, sopB* (also known as sigD), *sopD, sipB, pipA, pipB*, or *pipD* gene result in reduced fluid accumulation in ligated ileal loops (13, 22, 51, 52). However, there are several differences between *S. dublin* and *S. typhimurium* during infection of cattles. While cattles inoculated orally with *S. dublin* develop a systemic infection (41), *S. typhimurium* causes an infection which is primarily enteric, with only occasional bacteremia (53). Furthermore, 75% of *S. typhimurium* infections occur in calves less than 2 months of age (42). In contrast, *S. dublin* is associated at similar frequencies with morbidity in both young and adult calves (35, 42). Since differences between *S. dublin* and *S. typhimurium* infections are apparent, it is not clear whether mutations in orthologous genes result in identical virulence defects in both serotypes.

Although mutational inactivation of four genes, namely, *aroA, dap, galE*, and *invH*, has been shown to result in attenuation of *S. typhimurium* after oral infection of calves, evidence implicating other genes in *S. typhimurium* calf virulence is either indirect (based on data from bovine ligated ileal loops or competitive infection experiments) or has been inferred from studies performed with *S. dublin*. To obtain a more complete picture of the role in enteropathogenicity of virulence factors identified in various model systems, we have evaluated a collection of isogenic *S. typhimurium* mutants, including *S. dublin* invH mutants, for virulence during oral infection of calves. This comparative analysis provided an opportunity to assess the relative importance of virulence mechanisms during diarrheal disease caused by *S. typhimurium*. To facilitate comparison of our results with published data, the collection contained the well-characterized *S. typhimurium* *aroA* vaccine, which has been characterized previously for calf virulence. In addition, the collection contained strains with mutations in major *S. typhimurium* virulence determinants, including *Salmonella* pathogenicity island 1 (SPI-1), SPI-2, SPI-5, the LPS outer core region, and the *Salmonella* plasmid virulence (spv) operon.

The invasion-associated type III secretion system encoded by SPI-1 is required for entry of *S. typhimurium* into intestinal epithelial cells and for colonization of murine Peyer’s patches (49). Following secretion by the invasion-associated type III secretion system encoded by the invasion-associated type III secretion apparatus, was used to assess the contribution of SPI-2 to *S. typhimurium* calf virulence. Insertional inactivation of *spvB* renders *S. typhimurium* avirulent (more than 10,000-fold attenuated) for mice (32). A second virulence gene cluster required for bacterial growth in calves and spleens of mice, the *spv* operon, is encoded by the virulence plasmid of *S. typhimurium* (16). The first gene, *spvR*, encodes a transcriptional activator for the following four genes, *spvABCD*, whose functions are currently unknown (4). A Tn5 insertion in *spvR* results in a 1,000-fold-decreased colonization of murine splenic tissue by *S. typhimurium* (4), To assess the contribution of the *spv* operon to diarrheal disease, an *S. typhimurium* strain (STN272) carrying a transposon insertion in *spvR* was tested for virulence during oral infection of calves.

**Table 1. Derivatives of *S. typhimurium* ATCC 14028 used in this study**

<table>
<thead>
<tr>
<th>Strain</th>
<th>Genotype</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>IR715</td>
<td>14028 nalidixic acid resistant, wild type</td>
<td>47</td>
</tr>
<tr>
<td>STN61</td>
<td>IR715 hilA::mini-Tn5Km2</td>
<td>49</td>
</tr>
<tr>
<td>STN119</td>
<td>IR715 spB::mini-Tn5Km2</td>
<td>49</td>
</tr>
<tr>
<td>STN162</td>
<td>IR715 pgiH::mini-Tn5Km2</td>
<td>49</td>
</tr>
<tr>
<td>STN166</td>
<td>IR715 rfaL::mini-Tn5Km2</td>
<td>49</td>
</tr>
<tr>
<td>STN272</td>
<td>IR715 spvR::mini-Tn5Km2</td>
<td>49</td>
</tr>
<tr>
<td>SVM255</td>
<td>14028 sopB</td>
<td>18</td>
</tr>
<tr>
<td>BA1567</td>
<td>14028 sopB::MudJ</td>
<td>1</td>
</tr>
<tr>
<td>CL1509</td>
<td>14028 aroA::Tn10</td>
<td>48</td>
</tr>
</tbody>
</table>

**Materials and Methods**

**Bacterial strains and growth conditions.** Derivatives of *S. typhimurium* ATCC 14028, a bovine *S. typhimurium* isolate, are listed in Table 1. Strain IR715 is a spontaneous nalidixic acid-resistant derivative of ATCC 14028 (47), which is fully calf virulent (49). Bacteria were cultured aerobically at 37°C in Luria-Bertani (LB) broth or on LB agar (16 g/liter) plates. If appropriate, antibiotics were added at the indicated concentrations: kanamycin, 100 mg/liter; nalidixic acid, 50 mg/liter; tetracycline, 20 mg/liter.

**Experimental animals.** Milk-fed male Frisian-Holstein calves (*n = 42*), aged 3 to 4 weeks, were obtained from a commercial dairy calf rearing operation. The weight of the calves ranged between 45 and 52 kg. Animals were cared for according to American Association for Accreditation of Laboratory Animal Care guidelines. Calves were fed milk replacer twice daily and were given water ad libitum. Prior to their use for experiments, calves were evaluated for the absence of elevated leukocyte counts, fever, and fecal excretion of *Salmonella* serotypes. *Salmonella* scrofusae were detected in fecal swabs by enrichment in tetrathionate broth (Difco) and plating on brilliant green agar (BBL).

**Challenge.** Oral infection of the calves was performed as described previously (26). In brief, the optical density of overnight cultures at 600 nm was determined, and volumes containing approximately equal numbers of each strain were added to 50 ml of a suspension of 5% magnesium trisilicate, 5% sodium bicarbonate, 20 mg/liter.

**Examination of calves after challenge.** A previous report indicates that in calves which survive an *S. typhimurium* infection diarrhea does not persist beyond the 10th day postinfection (34). Calves were hence monitored for 10 days postinfection and then humanely euthanized. Weights and fecal scores of calves were recorded daily. Fecal score was used to assess the severity of diarrhea: 1 = normal feces; 2 = soft feces with loss of distinct conformation; 3 = diarrhea, loose feces with reduced solid matter; and 4 = diarrhea, aqueous feces with markedly reduced or little solid matter, blood, and shredds of fibrin. Shedding of *S. typhimurium* was monitored by taking daily fecal swabs, subsequently enriching them in tetrathionate broth (Difco), and plating them on brilliant green agar (BBL). When calves developed anorexia or were unable to stand, they were euthanized for humane reasons as described previously (26).

**Histopathology.** Euthanasia and preparation of tissue samples for histopathology were performed as described previously (26). At necropsy, tissue samples
from Peyer’s patches, ileum, and mesenteric lymph node were collected; a portion was homogenized in phosphate-buffered saline and plated in the presence of the appropriate antibiotics for enumeration of bacteria. The remaining tissue samples were coded for blind examination. Tissue samples were fixed in 10% formalin, embedded in paraffin, sectioned at 0.008 mm, and stained with hematoxylin and eosin for histopathological examination.

RESULTS

Mutations in *aroA* and SPI-1 reduce the severity of diarrhea in calves. Mutations in *sopB*, *prgH*, *hilA*, *rfaJ*, *spvR*, or *spiB* had previously been implicated in enteropathogenicity (1, 22, 26, 49, 52), but their effect on *S. typhimurium* virulence during oral infection of calves is not known. In this study, the virulence of a collection of *S. typhimurium* strains, each carrying a mutation in one of these genes, was compared to that of the isogenic wild type (IR715) during oral infection of calves. Two *S. typhimurium* *sopB* mutants were used in this study (Table 1). Strain SVM255 carries a 4-bp deletion within the *sopB* coding sequence, and this mutation is nonpolar on the expression of the downstream gene *pipC* (also known as *sigE*) (18). Strain BA1567 carries the transposon Mu3 inserted 100 bp downstream of the beginning of the *sopB* open reading frame (1). Strains STN61 and STN162 carry transposon insertions in the SPI-1 genes *hilA* (at bp 920 of the open reading frame) and *prgH* (at bp 129 of the open reading frame), respectively. The invasion-associated type III secretion system is nonfunctional in strains STN61 (*hilA*) and STN162 (*prgH*), as suggested by the reduced ability of these mutants to invade HEp-2 cells and to colonize murine Peyer’s patches (49). Strain STN166 is rough due to a mini-Tn5Km2 insertion at bp 564 of the *rfaJ* open reading frame (49). The *rfaJ* gene product is required for LPS outer core biosynthesis and mouse virulence (5, 27, 31). Strain STN272 carries a mini-Tn5Km2 insertion at bp 381 of the *spvR* open reading frame which renders this mutant defective for colonization of internal organs of mice (49). *SpvR* is a positive regulator of *spvABCD* expression and is required for full mouse virulence of *S. typhimurium* (4). Strain STN119 carries a transposon which is inserted at bp 909 of the *spiB* open reading frame (49). The *spiB* gene, which is part of the type III secretion apparatus encoded by SPI-2, is essential for *S. typhimurium* mouse virulence and growth within macrophages in vitro (32). Inactivation of *spiB* in strain STN119 results in a marked defect for colonization of internal organs of mice (49). Finally, an *S. typhimurium* *aroA* mutant (CL1509) was tested for calf virulence, since this mutation has previously been reported to result in avirulence and a reduced severity of diarrhea (46).

Calves infected with these *S. typhimurium* strains fell into two groups. Calves infected with strain IR715 (wild type), STN119 (*spiB*), STN166 (*rfaJ*), or STN272 (*spvR*) or either of two *sopB* mutants (SVM255 and BA1567) had aqueous diarrhea with feces containing various combinations of blood, fibrin, and mucus. These *S. typhimurium* strains could be detected in fecal swabs until day 9 postinfection unless lethal signs of disease developed earlier. The weight loss of animals infected at a dose of $10^{10}$ CFU/animal ranged between 1.5 and 5 kg, and terminal signs of illness, including anorexia and central nervous system depression, occurred between 1 and 3 days postinoculation.

In contrast, calves infected with strain CL1509 (*aroA*), STN61 (*hilA*), or STN162 (*prgH*) either had only soft feces for 3 to 4 days or had normal feces. Fecal shedding of these mutants ceased by day 6 postinfection in all but one calf. The weight of calves infected at a dose of $10^{10}$ CFU/animal remained constant or increased. After 10 days, all calves were healthy.

In conclusion, mutations in *hilA*, *prgH*, or *aroA* reduced the severity of diarrhea in calves, whereas animals infected with the wild type or strains carrying mutations in *spiB*, *rfaJ*, *spvR*, or *sopB* developed clinically severe diarrhea.

Mutations in *aroA* and SPI-1 result in a markedly reduced mortality in calves. The 50% lethal morbidity dose of *S. typhimurium* IR715 for calves is approximately $6 \times 10^{8}$ CFU/animal (49). Calves were infected orally at a dose of $10^{10}$ CFU/animal (Table 2). At this dose, all calves infected with IR715 (wild type), STN119 (*spiB*), STN272 (*spvR*), BA1567 (*sopB*), or SVM255 (*spvB*) developed signs of terminal illness and were euthanized. In contrast, all calves infected with STN61 (*hilA*), STN162 (*prgH*), or CL1509 (*aroA*) survived the infection and remained healthy. No mortality was observed when groups of two calves were infected with strain STN119 (*spiB*), STN272 (*spvR*), BA1567 (*sopB*), or SVM255 (*spvB*) (49) at a dose of $10^{8}$ CFU/animal, although fecal scores were similar at both doses. The wild type (*IR715*) caused lethal morbidity in six of eight calves infected orally with $10^{7}$ CFU/animal. In conclusion, mutations in *aroA*, *hilA*, and *prgH* attenuated *S. typhimurium* to a much greater degree than did mutations in *spiB*, *sopB*, or *spvR*.

Mutations in SPI-1, SPI-2, *rfaJ*, and *aroA* reduce the severity of intestinal lesions in calves. To more fully understand the disease processes and outcomes, it is necessary to study the development and spectrum of lesions in the target tissues. At necropsy, gross pathologic examination was performed with calves infected at a dose of $10^{10}$ CFU/animal and tissue samples from Peyer’s patches, ileum, and mesenteric lymph node were collected for histopathologic examination. The wild type, strain STN272 (*spvR*), and to a lesser extent strain SVM255 (*spvB*) produced macroscopically severe acute fibrinopurulent necrotizing enteritis with segmental or continuous pseudomembrane deposition (Fig. 1A) of the terminal 5 m of the ileum. Peyer’s patches of the middle to terminal ileum, and the cranial 1 to 2 m of the colon. Strains STN119 (*spiB*) and STN166 (*rfaJ*) produced lesions of moderate to marked subacute fibrinopurulent enteritis often confined to Peyer’s patches (Fig. 1B) of the terminal one-third of the ileum. No gross pathological lesions were detected during necropsy of calves infected with STN61 (*hilA*), STN162 (*prgH*), or CL1509 (*aroA*) (Fig. 1C).

Histopathologic analysis revealed severe lymphoid depletion and confirmed the severe acute fibrinopurulent necrotizing enteritis extending from the muscularis mucosa to the luminal

### Table 2: Virulence of *S. typhimurium* strains for 3- to 4-week-old Friesian-Holstein calves

<table>
<thead>
<tr>
<th>Strain</th>
<th>Genotype</th>
<th>No. dead/no. total</th>
<th>Avg time to death (days)</th>
<th>Avg fecal score</th>
</tr>
</thead>
<tbody>
<tr>
<td>IR715</td>
<td>Wild type</td>
<td>8/8</td>
<td>3.1</td>
<td>3.1</td>
</tr>
<tr>
<td>STN61</td>
<td><em>hilA</em></td>
<td>0/4</td>
<td>NA^b</td>
<td>1.9</td>
</tr>
<tr>
<td>STN119</td>
<td><em>spiB</em></td>
<td>2/2</td>
<td>2.3</td>
<td>3.8</td>
</tr>
<tr>
<td>STN162</td>
<td><em>prgH</em></td>
<td>0/2</td>
<td>NA</td>
<td>2.1</td>
</tr>
<tr>
<td>STN166</td>
<td><em>rfaJ</em></td>
<td>1/2</td>
<td>3.4</td>
<td></td>
</tr>
<tr>
<td>STN272</td>
<td><em>spvR</em></td>
<td>2/2</td>
<td>1.2</td>
<td>4</td>
</tr>
<tr>
<td>SVM255</td>
<td><em>sopB</em></td>
<td>2/2</td>
<td>4</td>
<td>3.8</td>
</tr>
<tr>
<td>BA1567</td>
<td><em>sopB</em></td>
<td>2/2</td>
<td>4</td>
<td>3.1</td>
</tr>
<tr>
<td>CL1509</td>
<td><em>aroA</em></td>
<td>0/2</td>
<td>NA</td>
<td>2.1</td>
</tr>
</tbody>
</table>

^a Fecal scores were used to assess the severity of diarrhea, where 1 = normal feces; 2 = soft feces with loss of distinct conformation; 3 = diarrhea, loose feces with reduced solid matter; and 4 = diarrhea, aqueous feces with markedly reduced or little solid matter, blood, or fibrin.

^b NA, not applicable.

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S. typhimurium hilA or prgH mutants, which caused only minor lesions (Fig. 1C and 2), have a more than 350-fold-reduced ability to colonize bovine Peyer’s patches (49). A mutation in rfaJ reduces the ability to colonize bovine Peyer’s patches 40-fold (49) and leads to lesions of intermediate severity (Fig. 1B and 2). An spvR mutant does not display a significant colonization defect for bovine Peyer’s patches (49) and causes severe lesions. However, an exception to this trend was seen with the spiB mutant (STN119), which has no significant colonization defect for bovine Peyer’s patches (49) and yet caused lesions of reduced severity (Fig. 1B).

\section*{DISCUSSION}

Virulence is a complex phenotype, which is only fully expressed during host-pathogen interaction in vivo. Therefore, the elucidation of mechanisms by which S. typhimurium produces diarrhea requires the use of an animal model, which resembles the natural course of infection and the typical signs of disease. The animal models commonly utilized to study virulence mechanisms of S. typhimurium are inbred mouse lineages, most frequently BALB/c. The disease produced by S. typhimurium in mice is similar to typhoid fever caused by Salmonella typhi in humans (33). However, mice do not develop diarrhea, and therefore, murine typhoid does not accurately model the enteritis caused by S. typhimurium in humans. Calves infected with S. typhimurium, on the other hand, develop acute enteritis and hence are better suited for the study of diarrheal disease (14). The high cost of the calf model precludes its use for screening a large number of putative virulence determinants. As a consequence, to date mutations in only four genes (aroA, invH, dap, and galE) have been shown to attenuate S. typhimurium during oral infection of cattle (6, 7, 46, 51). The goal of the present study was to extend this analysis to include other S. typhimurium genes, which have been suggested to be important for virulence in calves.

Recently, the invasion-associated type III secretion system located on SPI-1 has been implicated in enteropathogenicity of S. typhimurium and S. dublin. For instance, a mutation in invH, a gene located on SPI-1, causes attenuation during oral infection of calves (51). Inactivation of sopB, a SPI-1 gene encoding a type III secreted effector protein, reduces S. dublin-induced fluid secretion in bovine intestinal loops (13). Furthermore, a mutation in hilA, a positive regulator of genes located on SPI-1, reduces the ability of S. typhimurium to cause fluid accumulation in bovine ligated ileal loops (1) and to colonize intestinal tissue of calves during a competitive infection with the wild type (49). Similarly, an S. typhimurium prgH mutant is recovered in lower numbers than is the wild type from bovine intestinal tissue (49). Consistent with these reports, we found that mutations in hilA and prgH markedly reduced the severity of diarrhea during an S. typhimurium infection in calves (Table 2). Calves infected with STN61 (hilA) and STN162 (prgH) passed loose stools for 3 to 5 days and did not lose weight. While SPI-1-mediated colonization of intestinal tissues appears to be essential for bovine enteritis, it is less important during infection of mice. For instance, a mutation in hilA does not attenuate S. typhimurium ATCC 14028 during intragastric infection of mice (1) but caused marked attenuation (≥47-fold) after oral infection of calves (Table 2). Since bovine enteritis closely resembles S. typhimurium infections in humans, it is likely that virulence determinants important for
FIG. 2. Histopathology of Peyer’s patches and ileum from perinatal calves inoculated per os with $10^{10}$ CFU of the indicated strains of *S. typhimurium* per animal, photographed at a 40x magnification. Shown are hematoxylin-and-eosin-stained sections of Peyer’s patch (left column) and terminal ileum (right column). Short arrows indicate areas of marked lymphoid depletion; long arrows indicate zones of variable degrees of fibrinopurulent necrotizing ileitis at the mucosal surface.
enteropathogenicity in calves (e.g., SPI-1) are of greater importance for human disease than predicted by the mouse data.

In addition to SPI-1, SPI-5 has recently been implicated in calf enteropathogenicity of S. dublin. For example, S. dublin-mediated fluid secretion in bovine ligated ileal loops requires sopB (sigD), a gene located on SPI-5 which encodes a protein whose injection into epithelial cells is mediated by an SPI-1-dependent mechanism (13, 52). Surprisingly, calves inoculated with either S. typhimurium sopB mutant (SVM255 or BA1567) developed severe diarrhea, severe intestinal lesions, and lethal morbidity at wild-type levels (Table 2). A mutation in the S. dublin sopB gene causes a relatively weak reduction in fluid accumulation in bovine ligated ileal loops compared to that elicited by a sipB mutant (13) or a sopB sopD mutant (22). Hence, it is possible that the observation that an S. dublin sopB mutant elicits reduced fluid secretion in bovine intestinal segments while an S. typhimurium sopB mutant causes severe diarrhea during an oral infection may be explained by the differences in sensitivity of the loop assay and oral infection. Alternatively, the relative importance of sopB for S. dublin and S. typhimurium calf virulence may differ. Nevertheless, this result illustrates the importance of performing oral infection experiments to conclude that a mutation markedly reduces the virulence of S. typhimurium for calves.

Unlike S. dublin, which causes a systemic disease in calves, S. typhimurium produces primarily an enteric infection (41, 53). The virulence plasmid of S. dublin is not involved in the enteric phase of infection in calves but is required for full virulence during the early phase of infection in calves and is of greater importance in calves as live vaccines for calves. Am. J. Vet. Res. 48:504–510.


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