

A *Salmonella fim* Homologue in *Citrobacter freundii* Mediates Invasion In Vitro and Crossing of the Blood-Brain Barrier in the Rat Pup Model

Petra Hess,^{1†} Artur Altenhöfer,¹ A. Salam Khan,¹ Neda Daryab,^{1‡} Kwang Sik Kim,^{2§}
Jörg Hacker,¹ and Tobias A. Oelschlaeger^{1*}

Institut für Molekulare Infektionsbiologie, University of Würzburg, Würzburg, Germany,¹ and Division of Infectious Diseases, Children's Hospital of Los Angeles and University of Southern California, Los Angeles, California²

Received 16 September 2003/Returned for modification 13 November 2003/Accepted 18 May 2004

From the invasive *Citrobacter freundii* strain 3009, an invasion determinant was cloned, sequenced, and expressed. Sequence analysis of the determinant showed high homology with the *fim* determinant from *Salmonella enterica* serovar Typhimurium. The genes of the invasion determinant directed invasion of recombinant *Escherichia coli* K-12 strains into human epithelial cell lines of the bladder and gut as well as mannose-sensitive yeast agglutination and were termed *fim_{Cf}* genes. Expression of the Fim_{Cf} proteins was shown by ³⁵S labeling and/or Western blotting. In the infant rat model of experimental hematogenous meningitis, *C. freundii* strain 3009 and the in vitro invasive recombinant *E. coli* K-12 strain harboring the *fim_{Cf}* determinant reached the cerebrospinal fluid, in contrast to the case for the control strain. The *fim* determinant was also necessary for efficient in vitro invasion by *C. freundii*, because a deletion mutant was strongly reduced in its invasion efficiency. The mutation could be complemented in *trans* by the corresponding genes. Invasion by *C. freundii* could be blocked only by D-mannose, GlcNAc, and chitin hydrolysate and not by other carbohydrates tested. In contrast, yeast agglutination was not affected by GlcNAc or chitin hydrolysate. This finding indicated mannose residues to be essential for both yeast agglutination and invasion, whereas GlcNAc (oligomer) residues of host cells are involved exclusively in invasion. These results showed the *fim* determinant of *C. freundii* to be responsible for D-mannose- and GlcNAc-dependent in vitro invasion without being assembled into pili and for crossing of the blood-brain barrier in the infant rat model.

Citrobacter freundii strains are gram-negative, motile, rod-shaped bacteria of the family *Enterobacteriaceae*. They are widespread in nature and can be found in the environment in soil and water as well as in foodstuffs. *C. freundii* can be isolated from a wide variety of animals, such as household pets, birds, cattle, and fish. Although *C. freundii* is often considered a commensal of the human intestinal flora, this organism may cause urinary tract infections, diarrhea, gastritis, wound infections, and nosocomial infections such as pneumonia and, rarely, meningitis in newborns. Several probable virulence factors have been identified in *C. freundii*. Isolates from humans and beef samples were positive for the production of Shiga (-like) toxin II. This toxin is likely to be involved in enteropathogenicity caused by *C. freundii*. Furthermore, strains from patients with diarrhea produced a heat-stable enterotoxin identical to the 18-amino-acid *Escherichia coli* heat-stable enterotoxin STIIa. Another probable virulence factor is a capsule, which is closely related to the Vi capsule of *Salmonella enterica* serovar Typhi. Finally, the ability to invade several cell lines has been reported (22, 33).

Most invasive bacteria encode specific invasion systems. These invasion systems may be presented by a single surface protein such as the invasins of *Yersinia enterocolitica* or *Yersinia pseudotuberculosis* or the internalin A of *Listeria monocytogenes*. Other invasion systems are encoded by several genes and determine a type III secretion system and effector proteins injected into the host cell to be invaded. This kind of invasion system is employed by *Salmonella* and *Shigella*. In contrast, some invasive bacteria employ adhesins as invasins. Certain *E. coli* strains causing either intestinal or urinary tract infections harbor the *afa-3* adhesin gene cluster. One of the *afa-3* gene products, AfaD, is obviously not just an adhesin but also mediates invasion (23). Another nonfimbrial adhesin of pyelonephritis-associated *E. coli* is Dr-II which has been shown to direct internalization into HeLa cells (36). Related fimbrial adhesins inducing internalization are the Dr fimbriae of uropathogenic *E. coli* strains (16). Even certain variants of type 1 fimbriae are able to provoke bacterial invasion. These pili are the most widespread fimbrial adhesins among enterobacteria but vary in the amino acid sequence of the adhesive protein subunit FimH, located at the tip of the pilus (46). Some uropathogenic *E. coli* strains were reported to express type 1 pili which are not only essential for efficient infection of the urinary bladder but also responsible for the invasion of macrophages in the absence of opsonic antibodies and subsequent intracellular survival (2). Furthermore, these type 1 pili of uropathogenic *E. coli* are also able to induce bacterial internalization into urothelial cells of the bladder in a murine cystitis model (31).

Here, we report the molecular cloning, sequencing, and

* Corresponding author. Mailing address: Institut für Molekulare Infektionsbiologie, Röntgenring 11, 97070 Würzburg, Germany. Phone: 931 312150. Fax: 931 312578. E-mail: t.oelschlaeger@mail.uni-wuerzburg.de.

† Present address: MPI für Biochemie, Abteilung Strukturfor-

schung, 82152 Martinsried, Germany.

‡ Present address: Klinik und Poliklinik für Hautkrankheiten der Universität Würzburg, D-97080 Würzburg, Germany.

§ Present address: Pediatric Infectious Diseases, Johns Hopkins University School of Medicine, Baltimore, MD 21287.

TABLE 1. Bacterial strains and plasmids

Strain or plasmid	Characteristic(s) relevant for this study	Reference or source
Strains		
<i>C. freundii</i>		
3009	Human UTI isolate	33
3009-dz	$\Delta fim(D)HFZ_{CF}$, 3009 derivative	This study
<i>E. coli</i>		
AAEC189	K-12, Δfim , afimbriated	5
BL21(DE3)pLysS	B, F ⁻ <i>ompT hsdS_B(r_B⁻ m_B⁻) gal dcm</i> (DE3)(pLysS)(Cm ^r)	40
DH5 α	K-12, <i>fim</i> ⁺ , predominantly fimbriated	21
HB101	K-12, <i>fim</i> ⁺ , not fimbriated	10
<i>S. enterica</i> serovar Typhimurium C17		
	Clinical isolate	30
Plasmids		
pBR322'	2.7-kb BamHI/PvuII part of pBR322, Ap ^r	7
pJP5603	Suicide vector, Km ^r , <i>oriR6K</i> , <i>mobRP4</i>	35
pK19mob	Broad-host-range vector, <i>oriV oriT mobRP4</i> , Km ^r	39
pSU19	Cloning vector, Cm ^r	3
pT7-3	Expression vector, <i>bla</i> under $\phi 10$ promoter control, Ap ^r	41
pT7-6	Expression vector, <i>bla</i> not under $\phi 10$ promoter control, Ap ^r	41
pAA8	pSU19 carrying <i>fimAICDH_{CF}</i>	This study
pANN801-13	pBR322 carrying <i>sfaI</i> from <i>E. coli</i> strain 536, Ap ^r	20, 25
pAZZ50	pBR322 carrying <i>sfaII</i> from <i>E. coli</i> strain IHE3034, Ap ^r	18
pB7-3	<i>fimAICDHF_{CF}</i> under $\phi 10$ promoter control in pT7-3	This study
pB7-6	<i>fimAICDHF_{CF}</i> in pT7-6 antiparallel to the $\phi 10$ promoter	This study
pGB30	pHC79 carrying <i>fim_{Ec}</i> from <i>E. coli</i> strain 536, Ap ^r	19
pISF101	pACYC184 carrying <i>fim_{Sa}</i> from <i>S. enterica</i> serovar Typhimurium, Cm ^r	11
pMMP658-6	pBR322 carrying <i>sfr</i> from <i>E. coli</i> strain BK658, Ap ^r	34
pPH1	pSU19 carrying <i>fimAICDHF_{CF}</i>	This study
pPH4	Religated 7.1-kb HpaI/SnaI fragment of pPH1	This study
pPH13	4-kb Sall/KpnI of pPH4 in suicide vector pJP5603	This study
pPH19	6.2-kb PstI fragment of pTO3 in pBluescript II KS carrying <i>fimICDHF_{CF}</i>	This study
pPH23	pK19mob carrying <i>fimDHFZ_{CF}</i>	This study
pPH24	pPH1 with IS1 in <i>fimA_{CF}</i>	This study
pPIL110-54	pACYC184 carrying <i>foc</i> from <i>E. coli</i> strain AD110; Cm ^r	24, 44
pTO3	pBR322 carrying the whole <i>fim</i> cluster from <i>C. freundii</i>	22

analysis of an invasion determinant from a urinary tract *C. freundii* isolate. This invasion determinant shows high homology to the type 1 pilus determinant of *Salmonella*, which, in contrast, does not mediate invasion.

MATERIALS AND METHODS

Bacterial strains, growth conditions, and plasmids. The bacterial strains and plasmids used in this study are listed in Table 1. Recombinant bacteria were cultivated with shaking at 37°C overnight in Luria-Bertani (LB) broth supplemented with the appropriate antibiotic (i.e., 30 μ g of chloramphenicol per ml, 100 μ g of ampicillin per ml, or 20 μ g of kanamycin per ml). The antibiotics were purchased from Sigma (Deisenhofen, Germany). *C. freundii* strain 3009 (33) is from the strain collection of the Department of Bacterial Immunology, Walter Reed Institute of Research, Washington, D.C., and was grown in static nutrient broth at 37°C for 48 h. Before use in different experiments, *C. freundii* 3009 was usually passaged at least three times. M9 medium was supplemented with appropriate amino acids (0.1 mg/ml), 2 mM MgSO₄, 100 μ M CaCl₂, 0.2% glucose, and 60 μ M thiamine. Plasmid pTO3 was constructed by cloning partially digested (with Sau3A) chromosomal DNA of *C. freundii* 3009 into the BamHI site of cosmid vector pHC79. After transformation of noninvasive *E. coli* HB101, the resultant clones (3,840 clones) were screened for invasiveness in groups of 10 by performing the gentamicin protection assay (14). From the only group with a higher number of survivors than the average, each clone was tested for invasiveness individually. The only invasive clone was the one harboring cosmid pTO21052, which carries a ~50-kb chromosomal insert. After digestion of cosmid pTO21052 with PvuII, plasmid pTO3 was obtained, which consists of a 10.9-kb chromosomal insert and the 2.7-kb BamHI/PvuII part of cloning vector

pBR322 (22). Plasmid pPH1 was constructed by inserting the 9.6-kb EcoRI/SalI fragment of pTO3 into the EcoRI/SalI site of cloning vector pSU19. Ligating the 7.8-kb BamHI fragment of pTO3 to the pSU19 BamHI site resulted in a plasmid named pAA8. Plasmid pPH19 was constructed by cloning the 6.2-kb PstI fragment of pTO3 into the PstI site of cloning vector pBluescript II KS(+). By ligating the 9-kb EcoRI/XhoI insert of pTO3 to EcoRI- and Sall-digested pT7-3 and pT7-6, respectively, plasmids pB7-3 and pB7-6 were obtained. Plasmid pPH4 is a truncated derivative of pPH1 achieved by deletion of the HpaI/SnaI fragment, containing part of *fimD_{CF}* and genes *fimH_{CF}* to *fimZ_{CF}*, and subsequent religation. Suicide plasmid pPH13 was designed by ligating the 4-kb Sall/KpnI fragment from pPH4 into the multiple cloning site of suicide vector pJP5603 (35) which was digested with the same restriction enzymes. The deletion mutant *C. freundii* 3009-dz was obtained by transfer of the suicide plasmid pPH13 via conjugation from donor strain *E. coli* S17-1 λ pir to *C. freundii* strain 3009 and subsequent double crossover, which led to the exchange of the wild-type allele with the in vitro construct missing part of *fimD_{CF}* and genes *fimH_{CF}* to *fimZ_{CF}*. Plasmid pPH23 was constructed by inserting the 5.7-kb SmaI/SnaI fragment of pPH1 into the broad-host-range vector pK19mob (39) which was linearized with SmaI.

DNA sequencing and sequence alignments. Plasmid pTO3 was sequenced by the chain termination method (38), using a LI-COR DNA sequencer model 4000 (MWG-Biotech AG, Ebersberg, Germany). PCR was done with the Thermo Sequenase cycle sequencing kit (Amersham-Pharmacia, Freiburg, Germany) with primers complementary to the cloning vector pBR322 as well as forward and reverse primers designed from available DNA sequences (by primer walking). Sequence analysis was performed with the Genetics Computer Group and CLUSTALW programs and tools available at the website of the National Center for Biotechnology Information. Nucleotide and protein sequence homology

searches were done by BLAST search (National Center for Biotechnology Information).

Cell lines, media, and culture conditions. The human bladder epithelial cell line T24 was purchased from the American Type Culture Collection (Manassas, Va.). RT112, a human urinary bladder carcinoma cell line, was kindly provided by T. F. Meyer, Max-Planck-Institut für Infektionsbiologie, Berlin, Germany (8). The T24 cell line was cultivated in McCoy's 5A medium supplemented with 2 mM glutamine, nonessential amino acids, and 10% fetal calf serum. RT112 cells were grown in Waymouth MB 752/1 medium with 10% fetal calf serum. Both cell lines were cultivated in medium without antibiotics at 37°C in a 5% CO₂-95% air atmosphere with ~90% humidity and were split twice a week at a ratio of 1:5 to 1:10. All cell culture media and supplements were purchased from Gibco (Gaithersburg, Md.), except for McCoy's 5A, which was from C.C. Pro (Neustadt, Germany).

Invasion assay. For invasion assays, the human epithelial cells were seeded into 24-well plates (Falcon) and incubated in medium without antibiotics overnight at 37°C. Invasion assays were performed essentially as described by Elsinghorst (14). Briefly, a 5- to 50- μ l volume of a bacterial overnight culture was added to 2 ml of fresh LB medium and incubated with shaking until it reached the early logarithmic growth phase (optical density at 600 nm of 0.4 to 0.6). Approximately 1×10^5 to 2×10^6 bacteria were added to a confluent monolayer of epithelial cells and incubated for up to 3 h at 37°C in a 5% CO₂-95% air atmosphere (invasion period). The actual value for each inoculum was determined by a colony plate count. After the invasion period, the monolayer was washed twice with Earle's balanced salt solution, and fresh prewarmed medium containing 100 μ g of gentamicin per ml was added to kill the extracellular bacteria. After another 1 h incubation the monolayer was washed three times with Earle's balanced salts solution and lysed with 0.2% sodium deoxycholate in distilled water for 4 min. The viability of all strains used in this study was not affected by the 0.2% sodium deoxycholate treatment. The released intracellular bacteria were enumerated by a quantitative plate count. Invasion ability was expressed as the percentage of the inoculum surviving the gentamicin treatment. Each assay was conducted in duplicate and was independently repeated at least three times. Results are expressed as the means from all replicate experiments. In control experiments, the gentamicin sensitivity of all strains included in this study was demonstrated in the absence of epithelial cells by using equivalent bacterial numbers and under the same conditions as in invasion assays. All bacteria were killed in those control studies after treatment with gentamicin (100 μ g/ml) for 1 h. Epithelial cell viability and monolayer integrity were routinely monitored by addition of trypan blue (Hazleton Biologics, Lenexa, Kans.) and light microscopic analysis.

Invasion assays in the presence of carbohydrates. All of the carbohydrates used were directly dissolved in the appropriate cell culture medium at 100 mM, except for chitin hydrolysate (0.6 mg/ml). Chitin hydrolysate is a mixture of *N*-acetylglucosamine (GlcNAc) and oligomers of this carbohydrate (Vector Laboratories, Grünberg, Germany). In 24-well plates, the bacterial inoculum was added to 1 ml of cell culture medium with and without a particular carbohydrate and incubated with shaking at room temperature for 15 min. After removal of the tissue culture medium from the epithelial cell monolayer, the preincubated bacterial culture without and with carbohydrate was added to the monolayer and invasion assays were continued as described above. Carbohydrates were present during the invasion period. Inhibition of *C. freundii* 3009 internalization by chitin hydrolysate in a dose-dependent manner was analyzed at concentrations of 0.0375 to 0.6 mg/ml. Control studies under identical conditions but in the absence of human cells demonstrated that none of the carbohydrates used adversely affected bacterial viability. Trypan blue staining was performed to ensure human cell monolayer integrity under the assay conditions used.

Yeast cell agglutination. Prior to use, bacteria were routinely examined for type 1 fimbriae expression by mannose-sensitive yeast cell agglutination on glass slides (37). Type 1 fimbriae expression of bacteria was confirmed by agglutination after addition of an equal volume of baker's yeast suspension in phosphate-buffered saline (PBS) to the bacterial culture in the presence or absence of 2% D-mannose.

ELISA-based assay. In order to determine whether *sfaI*, *sfaII*, *sfr*, *foc*, and *fim* determinant-carrying recombinant bacteria and control strain HB101, harboring the plasmid vector pBR322, are fimbriated or not, enzyme-linked immunosorbent assays (ELISA) were performed (25). Overnight bacterial cultures were centrifuged, and bacterial pellets were resuspended in a carbonate buffer (pH 9.5) to a concentration of 10^9 CFU/ml. Flat-bottom 96-well ELISA plates (CML-CEB, Nemours, France) were coated with bacteria (200 μ l/well) and left overnight at 4°C. After removal of the unbound bacteria, the wells were blocked with 3% bovine serum albumin (BSA)-PBS (pH 7.4) for 2 h at 37°C and washed three times with PBS. Subsequently, 100 μ l of serially diluted fimbriae-specific rabbit

polyclonal antibody solutions in PBS-0.5% BSA were added to each well and incubated at 37°C for 90 min. After another washing step, horseradish peroxidase-conjugated goat anti-rabbit immunoglobulin G (Dako, Hamburg, Germany) in PBS-1% BSA (1:2,000) was added and incubated for 1 h at 37°C (100 μ l/well). Following a final wash, the bound enzyme was detected by the addition of substrate (100 μ l/well; Pierce ImmunoPure TMB substrate kit) for 5 to 30 min. The reaction was stopped by adding 100 μ l of 2 M H₂SO₄ per well. The *A*₄₅₀ was measured with an ELISA reader.

Adherence assay. Adherence was quantified by a modified invasion assay. For that assay, epithelial cells were seeded in wells of a 96-well plate and incubated for 24 h. To the confluent epithelial cell monolayer, 8 μ l (i.e., $\sim 10^8$ bacteria) of a static overnight culture of the strain of interest was added and incubated under cell culture conditions for 2 h. The number of epithelial cell-associated bacteria was determined by plate count. After five washing steps, cell-associated bacteria were resuspended in 0.2% Triton X-100 for 20 min, and 100- μ l volumes of appropriate dilutions were plated.

Heat extraction of fimbrial proteins. Type 1 fimbriae are heat extractable (26). For preparation of heat-extracted proteins, fimbriated bacteria were pelleted by centrifugation and suspended in 0.5 mM Tris-HCl (pH 7.4) containing 75 mM NaCl, followed by a 30-min incubation period at 60°C in a shaking water bath. After removal of bacteria by centrifugation, the crude fimbrial preparations were concentrated from the supernatant with a cellulose filter (Centricon YM-10; Millipore, Eschborn, Germany) by passing liquid and molecules smaller than 10 kDa through the filter.

Western blot analysis. Heat-extracted proteins were separated by electrophoresis (32 mA, 60 min) on a sodium dodecyl sulfate (SDS)-13% polyacrylamide gel (27) and were transferred to a nitrocellulose membrane by electrophoretic blotting (43). The blocked membranes were probed with various rabbit antisera. Following incubation of goat anti-rabbit immunoglobulin G conjugated with horseradish peroxidase (Dako), the membranes were developed by using an ECL kit (Amersham-Pharmacia) according to the manufacturer's instructions. Prestained full-range Rainbow marker (RPN 800; Amersham-Pharmacia) was used as a molecular weight standard. *C. freundii*-specific FimF and FimH polyclonal antisera were prepared by immunization of rabbits with synthesized peptides derived from FimF (LHSDSRTRLPLEQAS) and FimH (AGAGNRPEG INPQTK), respectively, conjugated to carrier molecule keyhole limpet hemocyanin (Eurogentec, Herstal, Belgium).

Autoradiography. To radiolabel Fim_{Cf} proteins, plasmids which carry a T7 promoter were used for specific *fim_{Cf}* gene expression with T7 RNA polymerase (41). Briefly, 30 μ l of BL21(pLys) (harboring various expression plasmids) overnight cultures was grown in 1 ml of LB medium supplemented with 1% glucose, 50 μ g of ampicillin per ml, and 20 μ g of chloramphenicol per ml, at 37°C with shaking, to an optical density at 600 nm of 0.5 to 0.7. The bacterial pellet was washed with LB medium and was resuspended in 1 ml of LB medium containing the above-mentioned antibiotics and 2 mM IPTG (isopropyl- β -D-thiogalactopyranoside). Following a 30-min incubation at 37°C with shaking, the bacteria were harvested, washed with M9 medium, and resolved in 1 ml of M9 medium without methionine and cysteine to be cultivated at 37°C for 1 h. Rifampin was added to a final concentration of 200 μ g/ml, and after another 25-min incubation, the plasmid proteins were labeled with a mixture of [³⁵S]methionine and [³⁵S]cysteine (10 μ Ci) (Pro-Mix; Amersham-Pharmacia). Whole-cell extracts obtained from bacteria were separated by electrophoresis on an SDS-13% polyacrylamide gel (27). After electrophoresis, the gel was stabilized by incubation for 30 min in a solution of 10% acetic acid and 10% methanol, followed by another 30-min incubation in a solution consisting of 10% glycerol, 10% methanol, and 1 M salicylic acid (pH 7.2). A PhosphorImager, kindly provided by J. Köhrle, University of Würzburg, was used to visualize the radiolabeled proteins.

Transmission electron microscopy. Bacteria from repeated subcultures in static liquid LB broth were resuspended in saline. The presence of type 1 pili and type 1 pilus-like adhesions on the surfaces of bacteria was detected by mannose-sensitive yeast agglutination. A 30- μ l aliquot of the bacterial suspension was placed on top of a Formvar-coated copper grid and left for 1 min. After sedimentation, the bacteria were stained with a 30- μ l drop of 2% uranyl acetate for 30 s. The grids were blotted dry and examined in a Zeiss 10A transmission electron microscope at 60 kV.

Neonatal rat model. The ability of *C. freundii* 3009, recombinant *E. coli* HB101(pPH1), and control strain HB101(pSU19) to cross the blood-brain barrier was examined as described by Wang et al. (45). Briefly, outbred, specific-pathogen-free Sprague-Dawley rats with timed conception were purchased from Charles River Breeding Laboratories (Wilmington, Mass.). The rats delivered in our vivarium 5 to 7 days after they arrived. At 5 days of age, all members of each litter were randomly divided into three groups to receive via intracardiac injection 3.9×10^7 , 8.2×10^8 , and 5.4×10^8 CFU of *C. freundii* 3009, recombinant

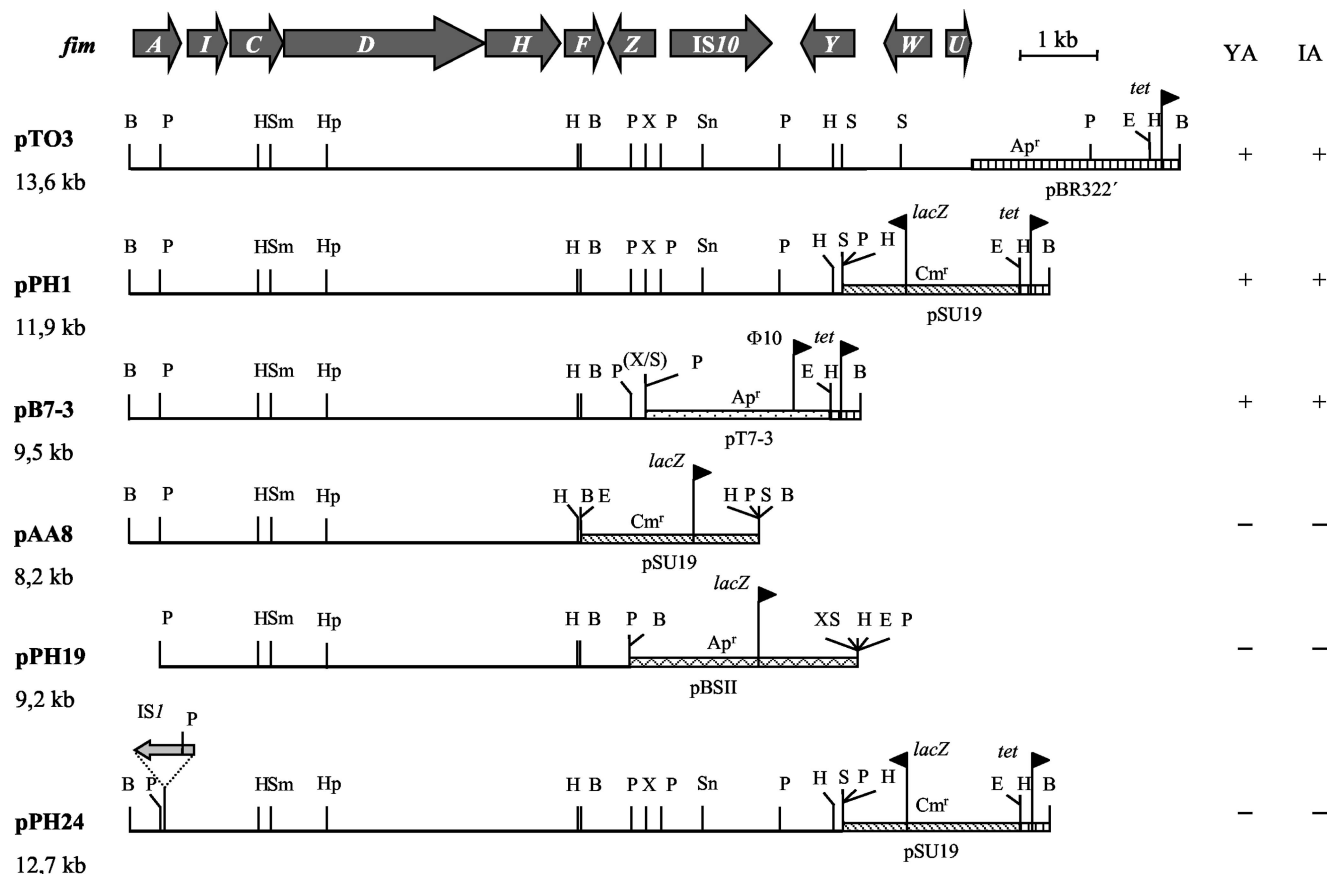


FIG. 1. Physical maps of plasmids containing the complete (pTO3) or parts of the type 1 fimbrial gene cluster from *C. freundii* strain 3009. The genes *fimA* to *fimF* on the plasmids are sufficient to enable recombinant *E. coli* HB101 or AAEC189 to agglutinate yeast cells and invade human bladder epithelial (T24) cells. B, BamHI; E, EcoRI; H, HindIII; Hp, HpaI; P, PstI; S, Sall; Sm, SmaI; Sn, SnaI; X, XhoI; YA, D-mannose-sensitive yeast agglutination; IA, invasion ability; ▨, pBR322-derived DNA; ▩, pSU19-derived DNA; ▧, pT7-3-derived DNA; ▤, pBluescript II KS-derived DNA.

E. coli HB101(pPH1), and *E. coli* HB101(pSU19), respectively. Approximately 1 to 2 h after bacterial inoculation, blood and cerebrospinal fluid specimens were obtained for quantitative cultures.

Nucleotide sequence accession number. The nucleotide sequence of the *C. freundii* *fim* gene cluster has been deposited in the GenBank database and given accession number AJ508060.

RESULTS

Cloning of an invasion determinant from *C. freundii* strain 3009. Recent studies revealed the ability of *C. freundii* strain 3009, a urinary tract isolate, to invade several human cell lines in vitro (33). To identify an invasion system in *C. freundii* strain 3009, cosmid cloning followed by screening for invasive clones was performed. To ensure cloning of large fragments of chromosomal *C. freundii* DNA partially digested with Sau3AI to the cosmid, cloning vector pHC79, previously linearized with BamHI, was chosen. Transformation of the noninvasive *E. coli* K-12 strain HB101 with the resulting recombinant cosmids was performed, and the transformants were screened for their ability to invade T24 human bladder epithelial cells. The invasion ability of recombinant clones was tested by employing the gentamicin protection assay. An invasive recombinant clone harboring cosmid pTO21502 with an insert of about 50 kb was identified. Digestion of pTO21052 with PvuII and religation

resulted in plasmid pTO3 (13.6 kb) (Fig. 1). This plasmid had lost the *cos* site of the vector pHC79, contained an insert of 10.9 kb, and mediated invasion into T24 cells (Fig. 1). Furthermore, the pTO3-carrying strain also invaded other human bladder cells, as well as epithelial cells of the gut and even human brain microvascular endothelial cells (Table 2).

Sequence analysis of the invasion determinant of *C. freundii* 3009. Sequencing of the insert of pTO3 revealed 11 open reading frames (Fig. 1). Of these, 10 were found to exhibit the highest rate of identity with the type 1 fimbrial gene cluster of *S. enterica* serovar Typhimurium (Fig. 2). In contrast to the fimbrial gene cluster of *S. enterica* serovar Typhimurium, an insertion sequence (IS) element identical to IS10 of *E. coli* was located between *fimZ* and *fimY* in *C. freundii* 3009. Except for the presence of IS10, the structure of the *C. freundii* invasion determinant was identical regarding number and arrangement to the *S. enterica* serovar Typhimurium *fim* genes (Fig. 2). In contrast, homology with the *fim* genes of *E. coli* K-12 was much lower. This is the case not only for the *E. coli* *fim* gene cluster (a paralogue) but also for the *E. coli* *sfm* gene cluster (an orthologue) (Fig. 2). In addition, the arrangements of genes in the gene cluster of *C. freundii* and the *fim* determinant of *E. coli* differed substantially (Fig. 2). Moreover, some genes

TABLE 2. In vitro invasion efficiencies for different cell lines by *C. freundii*, *S. enterica* serovar Typhimurium, and recombinant *E. coli* carrying various plasmids or vector controls

Cell line	Human origin	% Invasion ^a							
		<i>C. freundii</i>		<i>S. enterica</i> serovar Typhimurium C17	<i>E. coli</i> HB101 carrying:				
		3009	3009-dz		pTO3	pPH1	pB7-3	pSU19	pBR322
T24	Bladder	11.4 ± 6.6	2.2 ± 1.9	52.1 ± 23.0	9.7 ± 6.8	6.1 ± 2.2	5.3 ± 2.6	0.05 ± 0.04	0.01 ± 0.01
RT112	Bladder	27.3 ± 4.1	2.1 ± 0.8	ND ^b	9.9 ± 2.1	ND	ND	0.02 ± 0.01	0.03 ± 0.02
HCT8	Ileocecum	67.2 ± 29.6	ND	41.4 ± 39.3	3.7 ± 1.7	1.0 ± 0.6	3.3 ± 1.4	0.05 ± 0.04	0.01 ± 0.03
HBMEC ^c	Brain	3.3 ± 1.2	ND	ND	ND	2.6 ± 1.3	ND	ND	0.01 ± 0.01

^a Data are the means ± standard deviations from at least three independently performed invasion assays.

^b ND, not determined.

^c HBMEC, human brain microvascular endothelial cells.

are exclusively present either in the *C. freundii* gene cluster (e.g., the homologue of *fimY*) or in the *E. coli* *fim* determinant (e.g., *fimB* and *fimE*) (Fig. 2). The high homology of the cloned invasion genes of *C. freundii* as well as of the deduced amino acid sequence with the *fim* genes and the Fim proteins of *Salmonella* implied that this determinant represented a *fim* gene cluster. To further support this conclusion, yeast agglutination tests were performed with the recombinant *E. coli* strains HB101 and AAEC189 carrying plasmid pTO3. As is typical for Fim-mediated yeast agglutination, agglutination was

observed for both strains only in the absence of mannose. Therefore, the 10 genes with high homology to *S. enterica* serovar Typhimurium *fim* genes were termed *fim*_{CF} (Fig. 1). No agglutination at all was found for control strains carrying the plasmid vector pBR322.

Invasion efficiencies mediated by various adhesins. The high homology with *fim* genes of *Salmonella* and the property of mannose-sensitive yeast agglutination raised the question of whether other adhesin determinants might also mediate invasion in the experimental setting used. Therefore, invasion as-

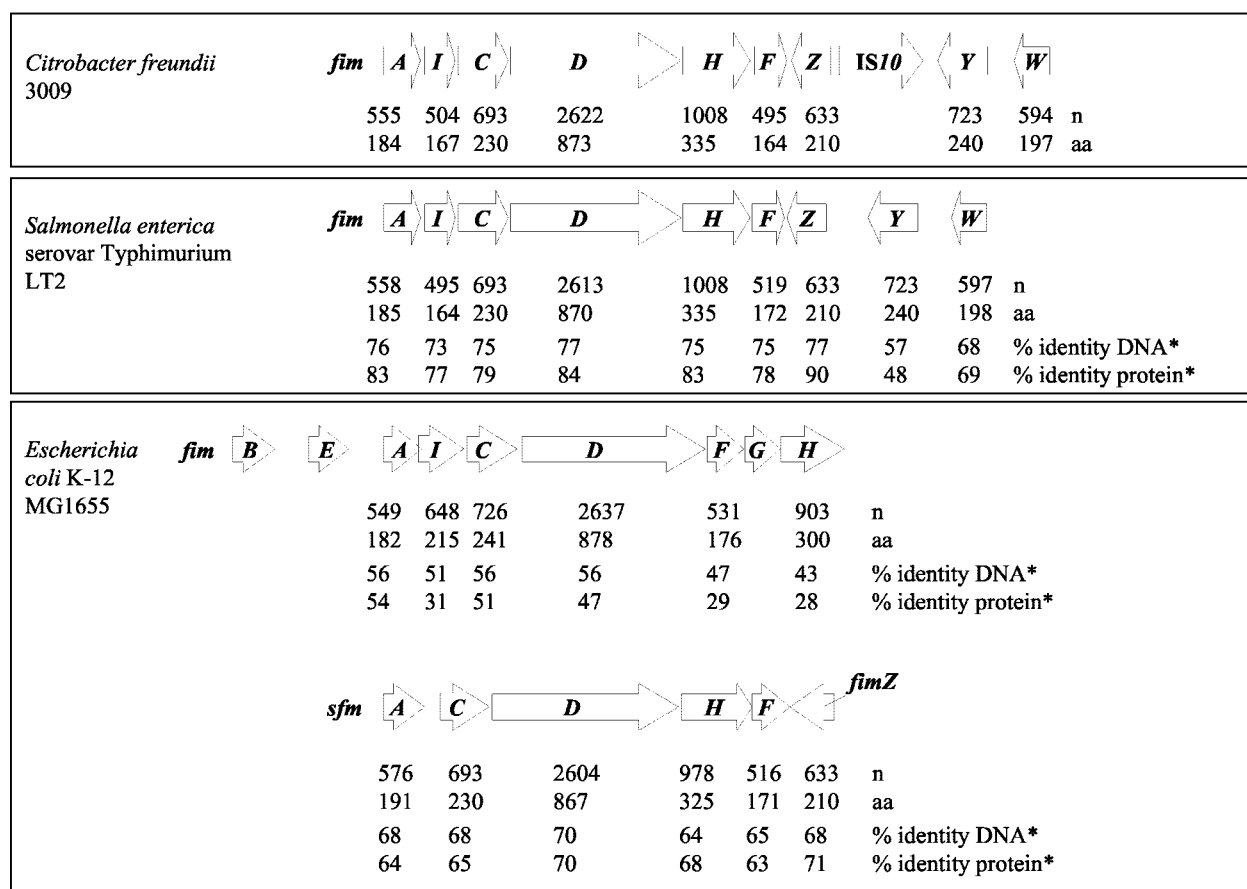


FIG. 2. Comparison of the genetic organization of the *fim* gene cluster from *C. freundii* strain 3009 to those from *S. enterica* serovar Typhimurium strain LT2 and *E. coli* K-12 strain MG1655, as well as to the *sfm* operon of *E. coli* MG1655 (an orthologue to the *S. enterica* serovar Typhimurium *fimACDHF* operon). The percent identity was determined by performing alignments by using CLUSTALW. n, number of nucleotides; aa, number of amino acids; *, percent identity either between *S. enterica* serovar Typhimurium and *C. freundii* or between *E. coli* and *C. freundii*.

TABLE 3. Internalization efficiencies of *C. freundii* strain 3009 and recombinant *E. coli* HB101 strains expressing different adhesin determinants with human bladder epithelial cell lines T24 and RT112

Plasmid or strain	Encoded adhesin ^a	Internalization efficiency ^b with:	
		T24 cells	RT112 cells
pBR322	None	0.020	0.0007
pPIL110-54	F1C	0.020	<0.0007
pMMP658-6	Sfr	0.007	<0.0007
pANN801-13	SfaI	0.010	<0.0007
pAZZ50	SfaII	0.050	0.0008
pGB30	Fim _{Ec}	0.020	<0.001
pISF101	Fim _{St}	0.002	ND ^c
pTO3	Fim _{Cf}	9.7	3.1
<i>C. freundii</i> 3009	Fim _{Cf}	11.4	27.3

^a Fim_{Ec}, type 1 pili from *E. coli*; Fim_{St}, type 1 pili from *S. enterica* serovar Typhimurium; Fim_{Cf}, type 1 pili from *C. freundii*.

^b The internalization efficiency is given as the percentage of the inoculum surviving gentamicin treatment. Data are the results of one of three independently performed assays with similar outcomes.

^c ND, not determined.

says were performed with recombinant *E. coli* HB101 strains each expressing one of various adhesins (Table 3). Adhesin expression was demonstrated for all adhesin determinants by ELISA (data not shown), and the adherence efficiency was quantified (Table 4). Although all adhesins were expressed and mediated adherence, not even the highly homologous *Salmonella fim* cluster was able to render the recombinant HB101 strain invasive. Only the cloned *C. freundii fim*_{Cf} determinant mediated invasion into T24 and RT112 human bladder epithelial cells (Table 3).

Determination of essential invasion genes. Sequence analysis of the *fim*_{Cf} gene cluster was the prerequisite for defined subcloning experiments and subsequent tests of the resulting transformants for invasiveness. Constructs no longer carrying *fimU*_{Cf}, *fimW*_{Cf}, and *fimY*_{Cf} (Fig. 1, pPH1) and in addition *fimZ*_{Cf} (Fig. 1, pB7-3) still mediated invasiveness and mannose-sensitive yeast agglutination (Fig. 1). The corresponding *fim* genes in *Salmonella* are known to have regulatory functions (42). Obviously, these *fim*_{Cf} genes are dispensable for the ex-

TABLE 4. Efficiencies of adherence of various recombinant *E. coli* strains and *C. freundii* to RT112 and T24 human bladder epithelial cells

Strain	Adhesin expressed	Adhering bacteria per well (10 ³) ^a with:	
		RT112 cells	T24 cells
<i>E. coli</i>			
HB101(pBR322)	None	12.0 ± 4.3	10.0 ± 2.8
HB101(pPIL110-54)	F1C	126.7 ± 41.1	210.0 ± 64.8
HB101(pMMP658-6)	Sfr	533.3 ± 124.7	531.3 ± 196.9
HB101(pANN801-13)	SfaI	416.7 ± 62.4	376.7 ± 33.0
HB101(pAZZ50)	SfaII	1020.0 ± 58.9	170.0 ± 21.6
HB101(pGB30)	Fim _{Ec}	493.3 ± 163.6	250.0 ± 40.8
HB101(pISF101)	Fim _{St}	160.0 ± 28.3	116.7 ± 17.1
HB101(pTO3)	Fim _{Cf}	60.7 ± 10.9	160.9 ± 65.6
<i>C. freundii</i> 3009	Fim _{Cf}	202.0 ± 20.4	333.3 ± 84.9

^a Data are means ± standard deviations.

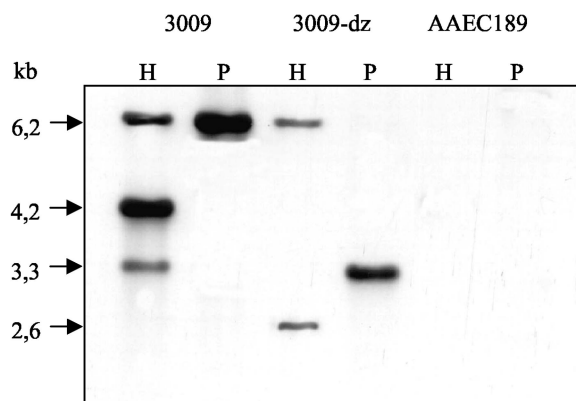


FIG. 3. Southern blot of HindIII (lanes H)- and PstI (lanes P)-digested chromosomal DNAs of wild-type strain *C. freundii* 3009, mutant 3009-dz (lacking part of *fimD*_{Cf} and genes *fimH*_{Cf} to *fimZ*_{Cf}), and control strain *E. coli* AAEC189 (in which the complete *fim*_{Ec} operon is deleted [5]). The 6.2-kb PstI fragment from plasmid pPH1 (Fig. 1) carrying *fimI*_{Cf} to *fimF*_{Cf} was used as a probe.

pression of adhesiveness and invasiveness. In contrast, further truncation of the insert of plasmid pB7-3 containing *fimA*_{Cf}, *fimI*_{Cf}, *fimC*_{Cf}, *fimD*_{Cf}, *fimH*_{Cf}, and *fimF*_{Cf} at either the *fimA*_{Cf} end (Fig. 1, pPH19) or the *fimF*_{Cf} end (Fig. 1, pAA8) resulted in the loss of adhesiveness and invasiveness (Fig. 1). Similarly, the insertion of an *IS1* element in *fimA*_{Cf} resulted in the loss of adhesiveness and invasiveness (Fig. 1). The conclusion was drawn from these results that genes *fimA*_{Cf} to *fimF*_{Cf} represented by the insert of plasmid pB7-3, were sufficient for mediating invasiveness and mannose-sensitive yeast agglutination. Inactivation of the *fim* determinant in *C. freundii* strain 3009 by deletion of the central part resulted in a mutant (3009-dz) lacking part of *fimD*_{Cf} and the genes *fimH*_{Cf}, *fimF*_{Cf}, and *fimZ*_{Cf}. The deletion was verified by PCR and Southern blotting (Fig. 3). The invasion efficiency in T24 cells of the mutant 3009-dz was reduced by 79% compared to that of the wild-type strain 3009. Introduction of plasmid pPH23, carrying *fimD*_{Cf}, *fimH*_{Cf}, *fimF*_{Cf}, and *fimZ*_{Cf} (Table 1), restored invasiveness to 62% of the wild-type level. Obviously, the *fim*_{Cf} determinant is essential for efficient invasiveness of *C. freundii* strain 3009.

In vivo studies. The ability of *C. freundii* 3009 to invade and replicate in human brain microvascular endothelial cells has been described previously (1). In addition, *C. freundii* is known to cause neonatal meningitis (12). We next examined the abilities of the wild-type *C. freundii* strain 3009 and the recombinant *E. coli* K-12 strain HB101(pPH1), harboring the *fim*_{Cf} genes, and the control strain HB101(pSU19) to penetrate into

TABLE 5. Effect of the *fim*_{Cf} determinant on the ability to reach the cerebrospinal fluid in the neonatal rat model

Strain	Bacteremia (log CFU/ml of blood) ^a	No. of animals with positive cerebrospinal fluid culture/total
<i>E. coli</i> HB101(pSU19)	5.31 ± 0.87	3/14 ^b
<i>E. coli</i> HB101(pPH1)	5.42 ± 0.71	14/20
<i>C. freundii</i> 3009	6.33 ± 1.02	7/10

^a Data are means ± standard deviations.

^b *P* < 0.05 by Fisher's exact test.

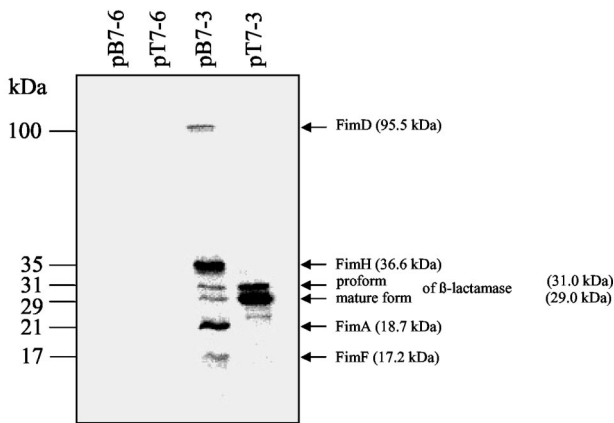


FIG. 4. Autoradiography after SDS-polyacrylamide gel electrophoresis of whole-cell lysates of *E. coli* BS21(pLysS) harboring recombinant plasmids. Proteins specifically expressed under control of the T7 promoter ϕ 10 were labeled with ^{35}S (see Materials and Methods) and separated on a 13% polyacrylamide gel by electrophoresis. The plasmids employed are indicated above the lanes. Molecular masses are indicated on the left. The most likely identities of the proteins, estimated by the molecular mass closest to the one deduced from the nucleotide sequence, are indicated at the right.

the cerebrospinal fluid in the newborn rat model (Table 5). As shown in Table 5, the magnitudes of bacteremia were similar in animals infected with HB101(pPH1) or HB101(pSU19). However, the occurrence of meningitis (defined as positive cerebrospinal fluid cultures) was significantly lower ($P < 0.05$) in animals receiving HB101(pSU19) (3 of 14; 21%) than in those receiving HB101(pPH1) (14 of 20; 70%).

Fim_{Cf} expression. The BamHI/XhoI fragment of pTO3 (Fig. 1) containing *fimA_{Cf}* to *fimF_{Cf}* was cloned into the expression vector pT7-3 under the control of the phage T7 promoter ϕ 10, resulting in plasmid pB7-3, and into pT7-6 antiparallel to the ϕ 10 promoter, creating pB7-6 (Fig. 1). These constructs as well as control plasmids were introduced into strain BL21DE3(pLysS). ^{35}S -labeled proteins were detected by autoradiography after separation by SDS-polyacrylamide gel electrophoresis. The molecular masses of the observed proteins were 100, 35, 31, 29, 21, and 17 kDa (Fig. 4). These values were in good agreement with the molecular masses calculated by employing SWISS-PROT (<http://www.expasy.ch/sprot>) for FimD_{Cf} (95.5 kDa), FimH_{Cf} (36.3 kDa), FimF_{Cf} (17.2 kDa), and the proform and the mature form of the β -lactamase encoded by pT7-3 (31 and 29 kDa), respectively. Only for the molecular mass of FimA_{Cf} determined by autoradiography (21 kDa) and deduced from the DNA sequence (18.7 kDa) was a discrepancy observed. Protein bands representing proteins with the molecular masses of FimI_{Cf} (18.1 kDa) and the chaperone FimC_{Cf} (25.1 kDa) could not be detected (Fig. 4).

In addition, FimF_{Cf} and FimH_{Cf} could be detected in Western blots with anti-FimF_{Cf} and anti-FimH_{Cf} rabbit sera in heat-extracted proteins of *E. coli* BL21(DE3)(pLysS)(pB7-3) and AAEC189(pPH1) (Fig. 5). These recombinant strains contained plasmids carrying *fimA_{Cf}* to *fimF_{Cf}* and *fimA_{Cf}* to *fimZ_{Cf}*, respectively (Fig. 1). No proteins were detected with the control strains BL21(DE3)(pLysS)(pT7-3) and AAEC189 with anti-FimH_{Cf} serum. However, the anti-FimH_{Cf} serum cross-reacted with FimH_{Ec} of the *E. coli* K-12 strain DH5 α due

to a sequence in FimH_{Ec} with 73% identity with the sequence of the peptide used to raise the antiserum (Fig. 5). The anti-FimF_{Cf} serum was not as specific, because it also recognized a few other protein bands representing proteins with molecular masses of >17 kDa from the control strains not expressing any Fim proteins (Fig. 5).

Detection of pili. In order to demonstrate the expression of pili by *C. freundii* strain 3009, specimens with this strain were negatively stained and inspected by transmission electron microscopy. The *C. freundii* culture as well as the other Fim-expressing strains used showed mannose-sensitive yeast agglutination, indicating the expression of the type 1 adhesin. The expression of other adhesins was demonstrated by ELISA and in adhesion assays (see Materials and Methods). Pili could be demonstrated for all of the recombinant *E. coli* HB101 strains tested, with one exception. *E. coli* strain HB101(pTO3), carrying the *fim* gene cluster from *C. freundii*, was bare; i.e., no pili could be detected at its surface. Similarly, *C. freundii* 3009 also showed no pili; however, flagella were clearly visible (Fig. 6).

Carbohydrate residues involved in *C. freundii* internalization and yeast agglutination. In general, certain carbohydrate residues on the surfaces of host cells function as adhesin receptors during the establishment of an infection. Besides mannose residues, which constitute the receptor structure for type 1 pili, several other carbohydrates were tested for interference with yeast agglutination and invasion of T24 cells. Invasion by *C. freundii* strain 3009 was inhibited by more than 80% only in the presence of 100 mM mannose, 100 mM GlcNAc, and 0.6 mg of chitin hydrolysate (i.e., a mixture of GlcNAc oligomers of various sizes) per ml. For chitin hydrolysate, dose-dependent inhibition of *Citrobacter* invasion was demonstrated (Fig. 7). No inhibition was observed in the presence of glucose, galactose, fucose, or *N*-acetylneuraminic acid (data not shown). The invasion efficiency of *S. enterica* serovar Typhimurium strain C17 was not affected by any of the carbohydrates employed (Fig. 7). Interestingly, yeast agglutination by *C. freundii*

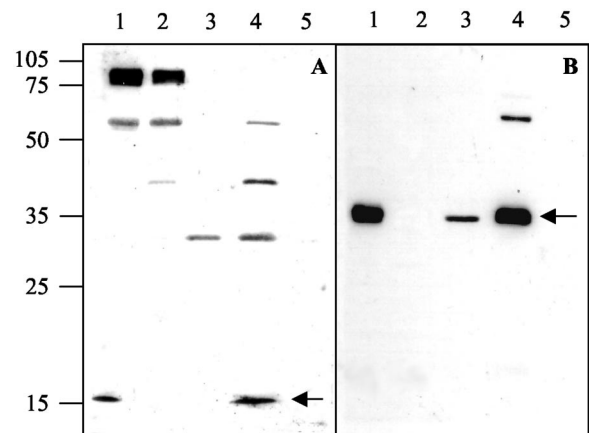


FIG. 5. Western blots of heat-extracted proteins transferred to a nitrocellulose membrane and probed with anti-FimF_{Cf} (A) and anti-FimH_{Cf} (B) sera. In panel A the protein band representing FimF (17 kDa) is indicated by an arrow. In panel B the arrow designates a polypeptide of about 36 kDa (FimH). Molecular masses of standard proteins, in kilodaltons, are indicated at the left. Lanes: 1, *E. coli* BL21(pLysS)(pB7-3); 2, *E. coli* BL21(pLysS)(pT7-3); 3, *E. coli* DH5 α ; 4, *E. coli* AAEC189(pPH1); 5, *E. coli* AAEC189.

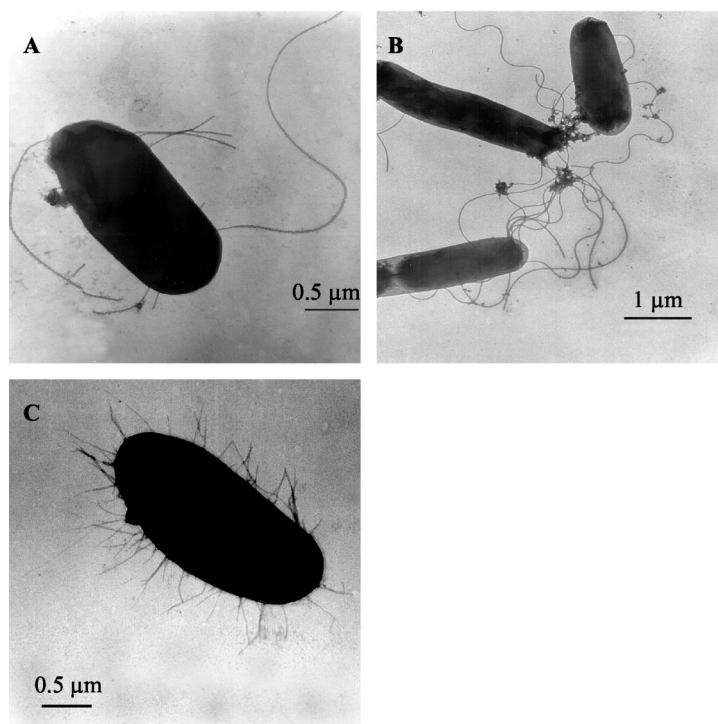


FIG. 6. Transmission electron microscopic images of *C. freundii* strain 3009 (A), *E. coli* strain AAEC189(pPH1) (B), and *E. coli* strain DH5α (C). The cultures of all strains used for the photographs presented here showed mannose-sensitive yeast agglutination.

strain 3009 was blocked only by mannose and not by any other carbohydrates tested.

DISCUSSION

In this study, we have identified and characterized an invasion determinant from *C. freundii* strain 3009 with high homology to the *Salmonella fim* gene cluster, encoding type 1 fimbriae. This was surprising, because *Citrobacter* is considered to be only distantly related to *Salmonella* (28). Although the *fim* determinant of *S. enterica* serovar Typhimurium is necessary for efficient invasion of HeLa cells, the corresponding type 1 pili are not sufficient to mediate invasion (4). This is in concordance with our observation that adherence mediated by neither the *fim* determinant from *E. coli* strain 536 carried by plasmid pGB30 nor that from *S. enterica* serovar Typhimurium strain LT2 resulted in internalization of bacteria. Deletion of the central part of the invasion determinant in *Citrobacter*, however, reduced invasiveness fivefold, whereas genes *fimA_{Cf}* to *fimF_{Cf}* imparted invasiveness to noninvasive *E. coli* K-12 strains. These data strongly implicate the Fim_{Cf} adhesin as the critical factor involved in uptake of *C. freundii* and recombinant *E. coli* strains by human T24 bladder epithelial cells.

As expected, the *fim_{Cf}* determinant was shown to be responsible for mannose-sensitive yeast agglutination. However, in spite of the high homology with the *Salmonella fim* gene cluster and the ability to mediate yeast agglutination, we were not able to demonstrate the presence of pili on *C. freundii* strain 3009 or any of the recombinant *E. coli* strains harboring the *fim_{Cf}* determinant. This resembles the situation of the Dr family of adhesins. This adhesin family consists of afimbrial and fimbrial

members with high amino acid sequence homology for genes A to D, encoding the chaperon and usher as well as the subunit proteins constituting the adhesin. The cause for the assembly of some of them into fimbriae while others are afimbrial adhesins is unclear (48).

The presence of a *fim* gene cluster with identical gene order and high homology at the nucleotide level to that of *Salmonella* might reflect horizontal transfer of this unit either from *Salmonella* to *Citrobacter* or vice versa, or *fim* may be ancestral to *Citrobacter*, *E. coli*, and *Salmonella* (9). If the first hypothesis is

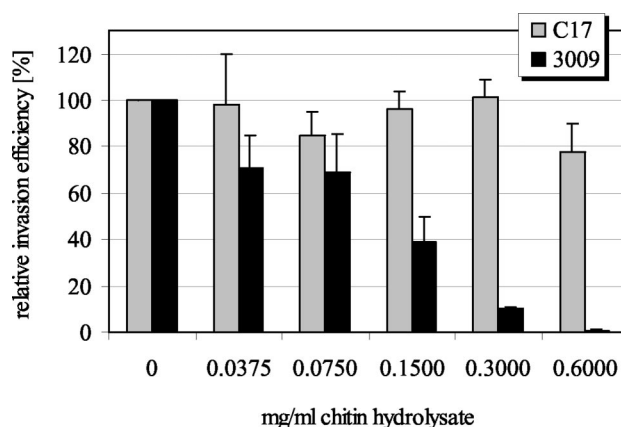


FIG. 7. Dose-dependent inhibition of *C. freundii* strain 3009 internalization into human bladder epithelial (T24) cells by chitin hydrolysate. *S. enterica* serovar Typhimurium strain C17 internalization was not inhibited. Results are presented as means \pm standard deviations.

correct, then after acquisition of the *fim* determinant by *Citrobacter*, it extended its function and the Fim subunit proteins were no longer assembled into a fimbrial structure. The new function was not just to direct adherence but also to direct invasion. Such a dual function of an adhesin, mediating adherence and invasion, is not a rare exception and is documented for a variety of adhesins, not just those of enterobacterial species (reviewed in reference 32).

The ability to invade host cells by expressing a Fim adhesin might well be employed by *Citrobacter* in vivo. As has been reported for uropathogenic *E. coli* in the mouse model, *C. freundii* might invade urothelial cells during a urinary tract infection (31). In addition, invasiveness could be used by *C. freundii* to initiate transcytosis and to cross the blood-brain barrier to cause neonatal meningitis. This can be hypothesized by taking into account that both *C. freundii* strain 3009 and a recombinant *E. coli* K-12 strain harboring the essential genes for invasiveness of the *fim*_{Cf} gene cluster were able to invade human epithelial and endothelial cells in vitro and crossed the blood-brain barrier in the rat pup model. These findings were also supported by a report by Badger et al. (1) demonstrating that *C. freundii* 3009 was able to invade, transcytose, and replicate inside human brain microvascular endothelial cells in vitro.

The mannose binding capacity of the FimH_{Cf} protein is essential for mediating efficient invasion by *C. freundii* as well as by the recombinant *E. coli* strains harboring the *fim*_{Cf} determinant. This was also reported for FimH_{Ec} of uropathogenic *E. coli* strain NU14, which is able to invade the human bladder epithelial cell line 5637 only in the absence of mannose (29). However, not every *fim*_{Ec} determinant is able to mediate invasion. The *fim*_{Ec} determinant, e.g., from *E. coli* 536 encoded by plasmid pGB30, did not mediate invasion. For *fimH*, single-nucleotide polymorphisms with adaptive advantage have been discovered in *E. coli* and *Salmonella enterica* serovar Typhimurium (6, 46). These polymorphisms are most likely responsible for the observed discrepancies between different *fim* determinants regarding the ability to mediate adherence with different efficiencies and for certain sequence variants in *E. coli* directing invasion (4, 47).

Besides mannose, there are several other carbohydrate residues that are frequently found as part of the glycocalyx of host cells, which might serve as receptors for bacterial adhesins. We tested fucose, galactose, GlcNAc, glucose, and *N*-acetylneuraminic acid for inhibitory effects on Fim_{Cf}-conducted invasion. None of the tested carbohydrates except GlcNAc showed an adverse effect. An anti-invasion effect for GlcNAc has also been reported for invasive *Klebsiella pneumoniae* strains (15). As for *Klebsiella*, a mixture of GlcNAc oligomers (i.e., chitin hydrolysate) produced a more pronounced inhibition of invasion than did GlcNAc monomers. In contrast to *C. freundii* invasion, *Klebsiella* invasion was not inhibited by mannose. There are no reports about the role of GlcNAc in invasion mediated by *E. coli* type 1 fimbriae. However, GlcNAc is the receptor structure recognized by several *E. coli* adhesins as F17 and K88 fimbriae (13, 17). Studies are in progress to clarify whether inhibition by GlcNAc of *C. freundii* invasion is due to a second binding capacity of FimH_{Cf} or whether another Fim_{Cf} subunit protein is involved in the invasion process by binding to GlcNAc.

ACKNOWLEDGMENTS

This work was supported in part by DFG grant Oe 135/3-2, grant SFB 479, and Graduiertenkolleg Infektiologie.

We are thankful to M. Keller (University of Bielefeld), S. P. Kidd (University of Birmingham), and A. Bäuml (Texas A&M University) for plasmids pK19mob, pJP5603, and pISF101, respectively. We thank J. Sheperd for help with sequence annotation, A. Reisenauer for support with invasion assays in the presence of carbohydrates, and C. A. Wass for technical assistance with the animal experiments.

REFERENCES

- Badger, J. L., M. F. Stins, and K. S. Kim. 1999. *Citrobacter freundii* invades and replicates in human brain microvascular endothelial cells. *Infect. Immun.* **67**:4208–4215.
- Baorto, D. M., Z. Gao, R. Malaviya, M. L. Dustin, A. van der Merwe, D. M. Lublin, and S. N. Abraham. 1997. Survival of FimH-expressing enterobacteria in macrophages relies on glycolipid traffic. *Nature* **389**:636–639.
- Bartolome, B., Y. Jubete, E. Martinez, and F. de la Cruz. 1991. Construction and properties of a family of pACYC184-derived cloning vectors compatible with pBR322 and its derivatives. *Gene* **102**:75–78.
- Bäumler, A. J., R. M. Tsois, and F. Heffron. 1996. Contribution of fimbrial operons to attachment to and invasion of epithelial cell lines by *Salmonella typhimurium*. *Infect. Immun.* **64**:1862–1865.
- Blomfield, I. C., M. S. McClain, and B. I. Eisenstein. 1991. Type 1 fimbriae mutants of *Escherichia coli* K-12: characterization of recognized afimbriate strains and construction of new *fim* deletion mutants. *Mol. Microbiol.* **5**:1439–1445.
- Boddicker, J. D., N. A. Ledebor, J. Jagnow, B. D. Jones, and S. Clegg. 2002. Differential binding to and biofilm formation on HEp-2 cells by *Salmonella enterica* serovar Typhimurium is dependent upon allelic variation in the *fimH* gene of the *fim* gene cluster. *Mol. Microbiol.* **45**:1255–1265.
- Bolivar, F., R. L. Rodriguez, P. J. Greene, M. C. Betlach, H. L. Heyneker, and H. W. Boyer. 1977. Construction and characterization of new cloning vehicles. II. A multipurpose cloning system. *Gene* **2**:95–113.
- Boxberger, H. J., and T. F. Meyer. 1994. A new method for the 3-D *in vitro* growth of human RT112 bladder carcinoma cells using the alginate culture technique. *Biol. Cell* **82**:109–119.
- Boyd, E. F., and D. L. Hartl. 1999. Analysis of the type 1 pilin gene cluster *fim* in *Salmonella*: its distinct evolutionary histories in the 5' and 3' regions. *J. Bacteriol.* **181**:1301–1308.
- Boyer, H. W., and D. Roulland-Dussoix. 1969. A complementation analysis of the restriction and modification of DNA in *Escherichia coli*. *J. Mol. Biol.* **41**:459–472.
- Clegg, S., B. K. Purcell, and J. Pruckler. 1987. Characterization of genes encoding type 1 fimbriae of *Klebsiella pneumoniae*, *Salmonella typhimurium*, and *Serratia marcescens*. *Infect. Immun.* **55**:281–287.
- Doran, T. I. 1999. The role of *Citrobacter* in clinical disease of children. *Clin. Infect. Dis.* **28**:384–394.
- el Mazouari, K., E. Oswald, J. P. Hernalsteens, P. Lintermans, and H. De Greve. 1994. F17-like fimbriae from an invasive *Escherichia coli* strain producing cytotoxic necrotizing factor type 2 toxin. *Infect. Immun.* **62**:2633–2638.
- Elsinghorst, E. A. 1994. Measurement of invasion by gentamicin resistance. *Methods Enzymol.* **236**:405–420.
- Fumagalli, O., B. D. Tall, C. Schipper, and T. A. Oelschlaeger. 1997. N-glycosylated proteins are involved in efficient internalization of *Klebsiella pneumoniae* by cultured human epithelial cells. *Infect. Immun.* **65**:4445–4451.
- Goluszko, P., V. Popov, R. Selvarangan, S. Nowicki, T. Pham, and B. J. Nowicki. 1997. Dr fimbriae operon of uropathogenic *Escherichia coli* mediates microtubule-dependent invasion to the HeLa epithelial cell line. *J. Infect. Dis.* **176**:158–167.
- Grange, P. A., M. A. Mouricout, S. B. Levery, D. H. Francis, and A. K. Erickson. 2002. Evaluation of receptor binding specificity of *Escherichia coli* K88 (F4) fimbrial adhesin variants using porcine serum transferrin and glycosphingolipids as model receptors. *Infect. Immun.* **70**:2336–2343.
- Hacker, J., H. Kestler, H. Hoshützky, K. Jann, F. Lottspeich, and T. K. Korhonen. 1993. Cloning and characterization of the S fimbrial adhesin II complex of an *Escherichia coli* O18:K1 meningitis isolate. *Infect. Immun.* **61**:544–550.
- Hacker, J., M. Ott, G. Blum, R. Marre, J. Heesemann, H. Tschäpe, and W. Goebel. 1992. Genetics of *Escherichia coli* uropathogenicity: analysis of the O6:K15:H31 isolate 536. *Zentbl. Bakteriol.* **276**:165–175.
- Hacker, J., G. Schmidt, C. Hughes, S. Knapp, M. Marget, and W. Goebel. 1985. Cloning and characterization of genes involved in production of mannose-resistant, neuraminidase-susceptible (X) fimbriae from a uropathogenic O6:K15:H31 *Escherichia coli* strain. *Infect. Immun.* **47**:434–440.
- Hanahan, D. 1983. Studies on transformation of *Escherichia coli* with plasmids. *J. Mol. Biol.* **166**:557–580.
- Hess, P., N. Daryab, K. Michaelis, A. Reisenauer, and T. A. Oelschlaeger. 2000. Type 1 pili of *Citrobacter freundii* mediate invasion into host cells. *Adv. Exp. Med. Biol.* **485**:225–235.

23. Jouve, M., M. I. Garcia, P. Courcoux, A. Labigne, P. Gounon, and C. Le Bouguenec. 1997. Adhesion to and invasion of HeLa cells by pathogenic *Escherichia coli* carrying the *afa-3* gene cluster are mediated by the AfaE and AfaD proteins, respectively. *Infect. Immun.* **65**:4082–4089.
24. Khan, A. S., B. Kniép, T. A. Oelschlaeger, I. Van Die, T. Korhonen, and J. Hacker. 2000. Receptor structure for F1C fimbriae of uropathogenic *Escherichia coli*. *Infect. Immun.* **68**:3541–3547.
25. Khan, A. S., I. Mühlendorfer, V. Demuth, U. Wallner, T. K. Korhonen, and J. Hacker. 2000. Functional analysis of the minor subunits of S fimbrial adhesion (SfaI) in pathogenic *Escherichia coli*. *Mol. Gen. Genet.* **263**:96–105.
26. Khan, A. S., and D. M. Schifferli. 1994. A minor 987P protein different from the structural fimbrial subunit is the adhesin. *Infect. Immun.* **62**:4233–4243.
27. Laemmli, U. K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* **227**:680–685.
28. Lawrence, J. G., D. L. Hartl, and H. Ochman. 1991. Molecular considerations in the evolution of bacterial genes. *J. Mol. Evol.* **33**:241–250.
29. Martinez, J. J., M. A. Mulvey, J. D. Schilling, J. S. Pinkner, and S. J. Hultgren. 2000. Type 1 pilus-mediated bacterial invasion of bladder epithelial cells. *EMBO J.* **19**:2803–2812.
30. Meier, C., T. A. Oelschlaeger, H. Merkert, T. K. Korhonen, and J. Hacker. 1996. Ability of *Escherichia coli* isolates that cause meningitis in newborns to invade epithelial and endothelial cells. *Infect. Immun.* **64**:2391–2399.
31. Mulvey, M. A., Y. S. Lopez-Boado, C. L. Wilson, R. Roth, W. C. Parks, J. Heuser, and S. J. Hultgren. 1998. Induction and evasion of host defenses by type 1-piliated uropathogenic *Escherichia coli*. *Science* **282**:1494–1497.
32. Oelschlaeger, T. A. 2001. Adhesins as invasins. *Int. J. Med. Microbiol.* **291**:7–14.
33. Oelschlaeger, T. A., P. Guerry, and D. J. Kopecko. 1993. Unusual microtubule-dependent endocytosis mechanisms triggered by *Campylobacter jejuni* and *Citrobacter freundii*. *Proc. Natl. Acad. Sci. USA* **90**:6884–6888.
34. Pawelzik, M., J. Heesemann, J. Hacker, and W. Opferkuch. 1988. Cloning and characterization of a new type of fimbria (S/F1C-related fimbria) expressed by an *Escherichia coli* O75:K1:H7 blood culture isolate. *Infect. Immun.* **56**:2918–2924.
35. Penfold, R. J., and J. M. Pemberton. 1992. An improved suicide vector for construction of chromosomal insertion mutations in bacteria. *Gene* **118**:145–146.
36. Pham, T. Q., P. Goluszko, V. Popov, S. Nowicki, and B. J. Nowicki. 1997. Molecular cloning and characterization of Dr-II, a nonfimbrial adhesin-I-like adhesin isolated from gestational pyelonephritis-associated *Escherichia coli* that binds to decay-accelerating factor. *Infect. Immun.* **65**:4309–4318.
37. Roe, A. J., C. Currie, D. G. E. Smith, and D. L. Gally. 2001. Analysis of type 1 fimbriae expression in verotoxigenic *Escherichia coli*: a comparison between serotypes O157 and O26. *Microbiology* **147**:145–152.
38. Sanger, F., S. Nicklen, and A. R. Coulson. 1977. DNA sequencing with chain-terminating inhibitors. *Proc. Natl. Acad. Sci. USA* **74**:5463–5467.
39. Schäfer, A., A. Tauch, W. Jäger, J. Kalinowski, G. Thierbach, and A. Pühler. 1994. Small mobilizable multi-purpose cloning vectors derived from the *Escherichia coli* plasmids pK18 and pK19: selection of defined deletions in the chromosome of *Corynebacterium glutamicum*. *Gene* **145**:69–73.
40. Studier, F. W. 1991. Use of bacteriophage T7 lysozyme to improve an inducible T7 expression system. *J. Mol. Biol.* **219**:37–44.
41. Tabor, S., and C. C. Richardson. 1985. A bacteriophage T7 RNA polymerase/promoter system for controlled exclusive expression of specific genes. *Proc. Natl. Acad. Sci. USA* **82**:1074–1078.
42. Tinker, J. K., and S. Clegg. 2001. Control of FimY translation and type 1 fimbrial production by the arginine tRNA encoded by *fimU* in *Salmonella enterica* serovar Typhimurium. *Mol. Microbiol.* **40**:757–768.
43. Towbin, H., T. Staehelin, and J. Gordon. 1979. Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications. *Proc. Natl. Acad. Sci. USA* **76**:4350–4354.
44. van Die, I., B. van Geffen, W. Hoekstra, and H. Bergmans. 1985. Type 1C fimbriae of a uropathogenic *Escherichia coli* strain: cloning and characterization of the genes involved in the expression of the 1C antigen and nucleotide sequence of the subunit gene. *Gene* **34**:187–196.
45. Wang, Y., S. H. Huang, C. A. Wass, M. F. Stins, and K. S. Kim. 1999. The gene locus *yijP* contributes to *Escherichia coli* K1 invasion of brain microvascular endothelial cells. *Infect. Immun.* **67**:4751–4756.
46. Weissman, S. J., S. L. Moseley, D. E. Dykhuizen, and E. V. Sokurenko. 2003. Enterobacterial adhesins and the case for studying SNPs in bacteria. *Trends Microbiol.* **11**:115–117.
47. Wilson, R. L., J. Elthon, S. Clegg, and B. D. Jones. 2000. *Salmonella enterica* serovars Gallinarum and Pullorum expressing *Salmonella enterica* serovar Typhimurium type 1 fimbriae exhibit increased invasiveness for mammalian cells. *Infect. Immun.* **68**:4782–4785.
48. Zhang, L., B. Foxman, P. Tallman, E. Cladera, C. Le Bouguenec, and C. F. Marrs. 1997. Distribution of *drb* genes coding for Dr binding adhesins among uropathogenic and fecal *Escherichia coli* isolates and identification of new subtypes. *Infect. Immun.* **65**:2011–2018.

Editor: J. T. Barbieri