



Assessment of the *Plasmodium falciparum* Preerythrocytic Antigen UIS3 as a Potential Candidate for a Malaria Vaccine

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ABSTRACT Efforts are under way to improve the efficacy of subunit malaria vaccines through assessments of new adjuvants, vaccination platforms, and antigens. In this study, we further assessed the *Plasmodium falciparum* antigen upregulated in infective sporozoites 3 (PfUIS3) as a vaccine candidate. PfUIS3 was expressed in the viral vectors chimpanzee adenovirus 63 (ChAd63) and modified vaccinia virus Ankara (MVA) and used to immunize mice in a prime-boost regimen. We previously demonstrated that this regimen could provide partial protection against challenge with chimeric *P. berghei* parasites expressing PfUIS3. We now show that ChAd63-MVA PfUIS3 can also provide partial cross-species protection against challenge with wild-type *P. berghei* parasites. We also show that PfUIS3-specific cellular memory responses could be recalled in human volunteers exposed to *P. falciparum* parasites in a controlled human malaria infection study. When ChAd63-MVA PfUIS3 was coadministered with the vaccine candidate *P. falciparum* thrombospondin-related adhesion protein (PfTRAP) expressed in the ChAd63-MVA system, there was no significant change in immunogenicity to either vaccine. However, when mice were challenged with double chimeric *P. berghei*-*P. falciparum* parasites expressing both PfUIS3 and PfTRAP, vaccine efficacy was improved to 100% sterile protection. This synergistic effect was evident only when the two vaccines were mixed and administered at the same site. We have therefore demonstrated that vaccination with PfUIS3 can induce a consistent delay in patent parasitemia across mouse strains and against chimeric parasites expressing PfUIS3 as well as wild-type *P. berghei*; when this vaccine is combined with another partially protective regimen (ChAd63-MVA PfTRAP), complete protection is induced.

KEYWORDS *Plasmodium*, UIS3, liver stage, malaria, sterile protection, synergy, vaccines

Plasmodium falciparum remains the leading causative agent of mortality due to human malaria, and eradication of this disease is a leading public health goal in many developing countries. Vaccination is considered to be a cost-effective preventative health tool and is considered vitally important for elimination of this disease (1). The leading malaria vaccine currently undergoing assessment in areas where malaria is endemic is RTS,S/AS01 (2), a subunit vaccine encoding the preerythrocytic antigen circumsporozoite protein (CSP). While this vaccine has proven to be partially effective (3), efforts continue to increase durable efficacy through assessments of new adjuvants, new delivery platforms, and/or new candidate antigens.

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Our past research demonstrated the capacity of viral vectors, as delivery platforms, to induce high-magnitude antigen-specific cellular immune responses in both animal models (4) and humans (5). Cellular immunity is essential for targeting the liver stage of the parasite's life cycle (6). A prime-boost regimen using the viral vectors chimpanzee adenovirus 63 (ChAd63) and modified vaccinia virus Ankara (MVA) has so far proved to be the most adept at inducing high-magnitude cellular immunity (7). Use of this regimen with vectors encoding the thrombospondin-related adhesion protein (TRAP) along with a multiepitope (ME) string resulted in moderate efficacy against *P. falciparum* sporozoites in malaria-naive adults (5) and in a field trial in a region where malaria is endemic (8). Building upon this work, we recently screened eight new virally vectored vaccines (containing *P. falciparum* preerythrocytic antigens as inserts) and compared their efficacies in mice against that induced by CSP or TRAP (9). We identified two antigens, *P. falciparum* liver-stage antigen 1 (PflSA1) and liver-stage-associated protein 2 (PflSAP2), that provided superior protection against challenge with chimeric *P. berghei*-*P. falciparum* parasites expressing the cognate *P. falciparum* antigen (9).

In addition to PflSA1 and PflSAP2, we observed that immunization of mice with viral vectors expressing the antigen upregulated in infective sporozoites 3 (UIS3) provided protection in BALB/c mice equal to that obtained by immunization with similar vectors expressing PfCSP. PfUIS3 did not induce protection in outbred CD-1 mice, but there was a median 1.1-day delay in the time to patent parasitemia (9). PfUIS3 is a 229-amino-acid protein that is a member of the early transcribed membrane protein (ETRAPM) family (10). PfUIS3 is hence also known as ETRAMP13 and is relatively conserved, with orthologs in *P. yoelii*, *P. vivax*, *P. cynomolgi*, *P. reichenowi*, *P. berghei*, and *P. chabaudi*, according to the PlasmoDB database (plasmodb.org). UIS3 became a protein of interest when it was shown to be upregulated in salivary gland sporozoites compared to oocyst sporozoites (PbUIS3) (11), and it has since been demonstrated that expression of this protein is restricted to sporozoites and liver-stage parasites for *P. yoelii* (12, 13), with no expression during the blood stage. Kaiser and colleagues (12) were also able to compare their PyUIS3 expression results with those of a *P. falciparum* microarray expression study (14), and they identified that PfUIS3 was also highly upregulated in sporozoites compared to asexual blood-stage parasites. This confirms an earlier report that PfUIS3 is not expressed during blood stages (10). UIS3 was subsequently shown to be essential for early liver-stage development in *P. berghei* (15) and *P. yoelii* (16). Parasites without UIS3 can still invade liver cells but fail to develop into mature liver-stage schizonts and fail to reach the blood stage. PfUIS3 is predicted to have an N-terminal signal peptide and two transmembrane domains (the first overlapping the predicted signal peptide) (see Fig. S1 in the supplemental material), suggesting that it may be localized to the membrane. Indeed, evidence of localization to the parasitophorous vacuole membrane (PVM) has been demonstrated for PyUIS3 (17). While the function of UIS3 is still unclear, the protein likely has a role in the importation of fatty acids into the PVM (17, 18). Utilizing PyUIS3 in a mouse liver model, it was shown that this protein directly interacts with the liver-fatty acid binding protein (L-FABP). L-FABPs have a known role in facilitating the utilization of fatty acids in liver cells (19), and it was demonstrated that levels of L-FABP in host hepatocytes directly correlated with liver-stage parasite growth (17). There is also evidence that PfUIS3 interacts directly with human L-FABP (18), supporting this proposed function. In addition to our recent work, PyUIS3 has been shown to provide protection against malaria when administered in combination with PyFalstatin. Using a vaccination regimen of DNA followed by a replication-competent vaccinia virus vector, these antigens together resulted in 43% sterile protection in outbred mice (13).

In the current study, we aimed to further investigate the potential of PfUIS3 as a vaccine candidate. We demonstrate here that CD8⁺ T cells are critical for protection induced by ChAd63-MVA PfUIS3, providing further evidence that this vaccine targets PfUIS3 expressed in liver-stage parasites. We also show that PfUIS3 is capable of inducing protection not only against chimeric *P. berghei*-*P. falciparum* parasites expressing PfUIS3 but also against wild-type (WT) *P. berghei* parasites. In addition, we com-

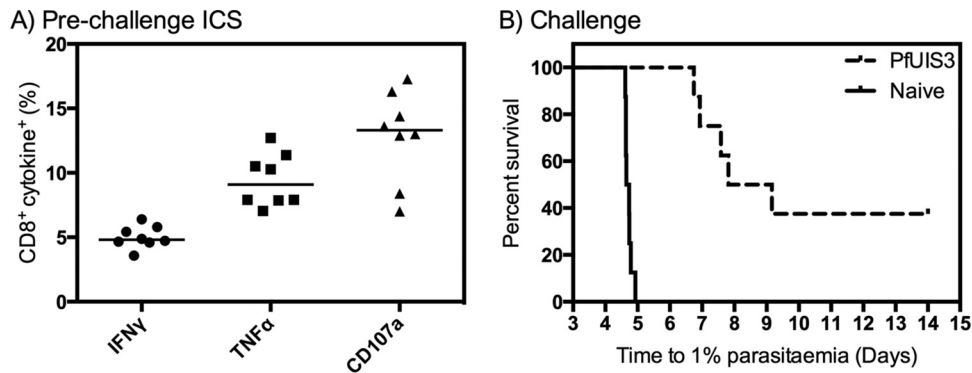


FIG 1 PfUIS3 protects C57BL/6 mice against chimeric *P. berghei*-*P. falciparum* challenge. C57BL/6 mice ($n = 8$) were vaccinated i.m. with 1×10^8 inclusion-forming units (IFU) of ChAd63-PfUIS3 followed 8 weeks later by 6×10^6 PFU of MVA-PfUIS3. (A) Cellular immunogenicity was assessed in the blood at 7 days postboost by ICS after stimulation for 6 h with an overlapping peptide pool. Both medians and individual data points are shown. (B) At 10 days postboost, mice were challenged i.v. with 1,000 chimeric *P. berghei*-*P. falciparum* sporozoites expressing PfUIS3, as were eight naive control mice. The log-rank (Mantel-Cox) test was used to assess the difference between the survival curves ($P < 0.0001$).

pared cellular and humoral immune responses of mice immunized with PfUIS3 either alone or in combination with PfTRAP, and we found no significant differences. Furthermore, when both PfUIS3 and PfTRAP were coadministered as a mixture, we observed 100% sterile protection against challenge with double chimeric *P. berghei*-*P. falciparum* parasites expressing both PfUIS3 and PfTRAP. We also showed that PfUIS3 memory responses could be recalled in human volunteers recently exposed to *P. falciparum* parasites. Together, our results suggest that PfUIS3 may be considered for inclusion in a multiantigen subunit vaccine.

RESULTS

PfUIS3 can provide protection against challenge with chimeric *P. berghei*-*P. falciparum* parasites in multiple strains of mice. We previously demonstrated that PfUIS3, administered in a ChAd63-MVA prime-boost regimen with an 8-week interval, could induce protective immune responses in BALB/c mice as determined by a significant delay in the time to 1% parasitemia after challenge with chimeric *P. berghei*-*P. falciparum* parasites expressing PfUIS3 (9). We also identified conserved immunogenic epitopes for PfUIS3 in both BALB/c and C57BL/6 mice (20). Therefore, we initially wished to determine whether PfUIS3 could induce protective immunity in C57BL/6 mice, an additional strain of inbred mice. C57BL/6 mice were vaccinated with ChAd63-MVA PfUIS3, using an 8-week interval, followed 10 days later by challenge with 1,000 chimeric *P. berghei*-*P. falciparum* sporozoites injected intravenously (i.v.). Vaccination of C57BL/6 mice induced a strong proinflammatory response in the blood, with a median of 4.8% CD8⁺ IFN- γ ⁺ cells (Fig. 1A), and provided a significant delay in the time to 1% parasitemia while also sterilely protecting 3/8 mice ($P < 0.0001$ by the log-rank [Mantel-Cox] test) (Fig. 1B). This experiment was repeated, and again, PfUIS3 vaccination provided a significant delay in the time to 1% parasitemia, and it sterilely protected 2/8 mice ($P < 0.0001$) (data not shown).

PfUIS3 can provide protection against WT *P. berghei* challenge. The UIS3 gene is present in both the *P. falciparum* and *P. berghei* genomes. Using the European Bioinformatics Institute EMBOSS Needle pairwise sequence alignment tool (http://www.ebi.ac.uk/Tools/psa/emboss_needle/), we found 54% sequence similarity and 33% identity between the *P. falciparum* and *P. berghei* UIS3 sequences. Given that previous research demonstrated the feasibility of cross-species protection using subunit vaccines (21, 22), we next assessed whether PfUIS3 could provide protection of BALB/c mice against challenge with WT *P. berghei* sporozoites. A moderate immune response was detected in BALB/c mice, as expected (9), with a median of 9.0% CD8⁺ IFN- γ ⁺ cells in the blood (Fig. 2A). Upon challenge with 1,000 WT *P. berghei* sporozoites

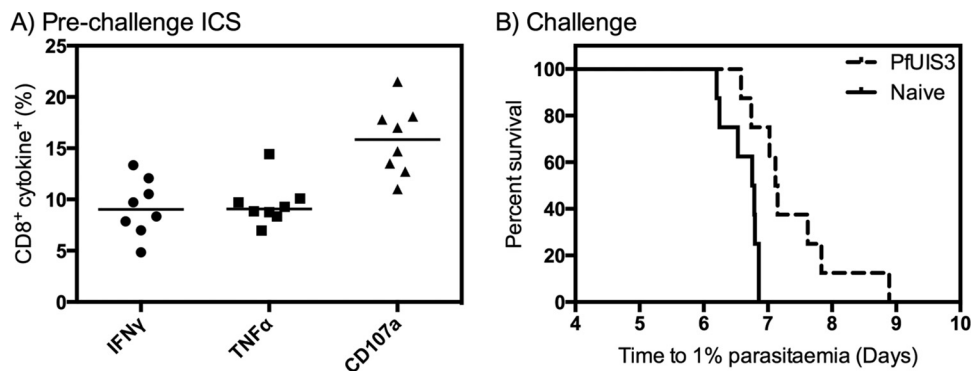


FIG 2 PfUIS3 protects BALB/c mice against WT *P. berghei* challenge. BALB/c mice ($n = 8$) were vaccinated i.m. with 1×10^8 IFU of ChAd63-PfUIS3 followed 8 weeks later by 1×10^7 PFU of MVA-PfUIS3. (A) Cellular immunogenicity was assessed in the blood at 6 days postboost by ICS after stimulation for 6 h with an overlapping peptide pool. Both medians and individual data points are shown. (B) At 8 days postboost, mice were challenged i.v. with 1,000 WT *P. berghei* sporozoites, as were eight naive control mice. The log-rank (Mantel-Cox) test was used to assess the difference between the survival curves ($P = 0.0048$).

at 8 days post-MVA boost, there was a statistically significant delay in the time to reach 1% parasitemia in vaccinated mice compared to naive controls (median delay, 0.36 day; $P = 0.0048$ by the log-rank [Mantel-Cox] test) (Fig. 2B). No mice were sterilely protected. In comparison, no protection against WT *P. berghei* challenge was observed in mice that were immunized with either of two other candidate antigens with similar levels of sequence similarity and identity between the *P. falciparum* and *P. berghei* orthologs (CelTOS, 65% and 45%, respectively; and Falstatin, 53% and 35%, respectively) (see Fig. S6 in the supplemental material).

PfUIS3 likely provides protection through antigen-specific CD8⁺ T cells. Because UIS3 is expressed during both the sporozoite and liver stages (15, 17) and vaccination induces both cellular and humoral immune responses (9), we next wished to determine the mechanism of protection observed in BALB/c mice. We depleted either CD4⁺ or CD8⁺ T cells from PfUIS3-vaccinated BALB/c mice prior to challenge with 1,000 chimeric *P. berghei*-*P. falciparum* parasites expressing PfUIS3. Depletion of CD8⁺ T cells completely abolished the protective effect of the vaccine, with no significant difference in the times to reach 1% parasitemia between this group of mice and naive controls and a significant difference compared to mice injected with an IgG control ($P < 0.0001$ by the log-rank [Mantel-Cox] test) (Fig. 3A). The median time to 1% parasitemia was reduced from 7.37 days for the IgG control mice to 5.57 days for CD8⁺ cell-depleted mice. CD4⁺ depletion also significantly reduced the efficacy compared to that with IgG control mice ($P = 0.0044$); however, this regimen still provided some degree of protection compared to naive mice ($P < 0.0001$). We previously identified a conserved epitope in BALB/c, C57BL/6, and HLA-A2-transgenic mice that elicited a CD4⁺ T cell response (20); this may be an important feature of PfUIS3 for the consistency in protection across mouse strains that we observed. We confirmed that either CD4⁺ or CD8⁺ T cells remained depleted 4 days following challenge (Fig. 3B).

We also observed PfUIS3-specific CD8⁺ T cells in the livers of vaccinated BALB/c and C57BL/6 mice, with a magnitude equal to or greater than that observed in the spleen (Fig. 4A). The majority of PfUIS3-specific cells (defined by gamma interferon [IFN- γ] secretion) in the liver were of the effector memory subtype (CD62L⁻ CD127⁺) (Fig. 4B). For both BALB/c and C57BL/6 mice, a significantly larger proportion of antigen-specific CD8⁺ cells were of the effector memory subset in the liver than in the spleen ($P < 0.0001$ and $P = 0.0027$, respectively, by two-way analysis of variance [ANOVA]).

Coadministration of ChAd63-MVA PfUIS3 with the vaccine ChAd63-MVA ME-TRAP results in 100% sterile protection in BALB/c mice. ME-TRAP is one of the leading preerythrocytic *P. falciparum* vaccine candidates (5). We wanted to determine if there was any impact on immunogenicity when PfUIS3 was coadministered with a PfTRAP-based vaccine. As described in Materials and Methods, we chose to use the

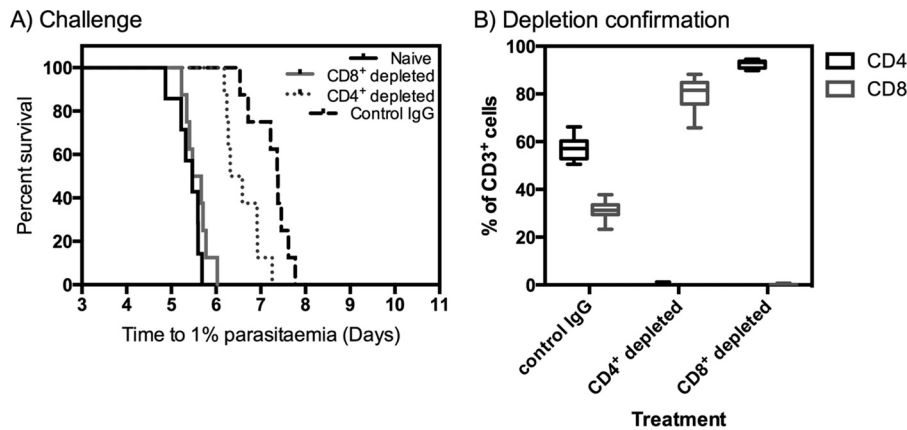


FIG 3 PfUIS3 requires CD8⁺ T cells to provide protection against chimeric *P. berghei*-*P. falciparum* challenge in BALB/c mice. BALB/c mice ($n = 3$ groups of 8) were vaccinated i.m. with 1×10^8 IFU of ChAd63-PfUIS3 followed 8 weeks later by 5×10^6 PFU of MVA-PfUIS3. Mice were then injected intraperitoneally (i.p.) with 100 μ g of MAb to either CD4⁺ (GK1.5) or CD8⁺ (8.43) or with an IgG MAb control at days 8, 9, and 10 postboost. (A) At day 10, all mice were challenged i.v. with 1,000 chimeric *P. berghei*-*P. falciparum* sporozoites expressing PfUIS3, as were seven naive control mice. The log-rank (Mantel-Cox) test was used to assess differences between the survival curves: for CD8⁺ cell-depleted versus naive mice, $P = 0.13$; for CD8⁺ cell-depleted versus control IgG-treated mice, $P < 0.0001$; for CD4⁺ cell-depleted versus naive mice, $P < 0.0001$; and for CD4⁺ cell-depleted versus IgG control-treated mice, $P = 0.0044$. (B) Depletion of CD8⁺ or CD4⁺ T cells was confirmed in the blood at day 4 with respect to challenge. Cells were stained with anti-CD4-FITC clone RM4-5, 1/200 anti-CD8-PerCP-Cy5.5 clone 53-6.7, and 1/50 anti-CD3 ϵ -APC, and the proportion of CD3⁺ cells that were CD4⁺ or CD8⁺ is shown.

vaccine PfTRIP, as this uses the TRAP sequence of *P. falciparum* strain 3D7, the same strain used for the UIS3 sequence. We observed no statistically significant differences in cellular or humoral immune response as measured by blood intracellular cytokine staining (ICS) or luminescence immunoprecipitation system (LIPS) assay between BALB/c mice immunized with the two vaccines administered together and mice immunized with either vaccine alone (Fig. 5). There was a trend toward a lower PfTRAP-specific CD4⁺ IFN- γ ⁺ response when the PfTRIP vaccine was coadministered with PfUIS3 (mixed); however, this was not statistically significant. Conversely, coadministration (whether at separate sites or mixed) resulted in slightly higher TRAP-specific IgG antibody responses than those obtained with PfTRIP administered alone (Fig. 5A). There

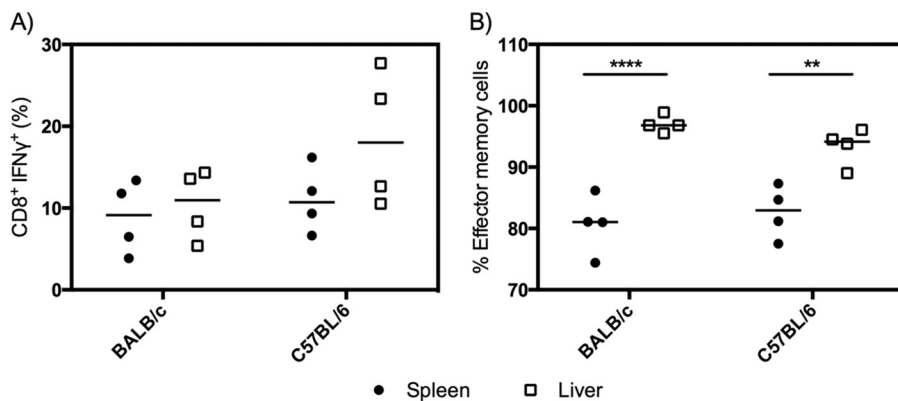


FIG 4 PfUIS3 vaccination induces antigen-specific effector memory cells in the livers of both BALB/c and C57BL/c mice. BALB/c or C57BL/6 mice ($n = 4$ per strain) were vaccinated i.m. with 1×10^8 IFU of ChAd63-PfUIS3 followed 8 weeks later by 1×10^6 PFU of MVA-PfUIS3. Cellular immunogenicity was assessed in both the spleen and liver at 2 weeks postboost. (A) Percentages of CD8⁺ cells secreting IFN- γ in each organ. Closed circles, spleen; open squares, liver. (B) Percentages of antigen-specific cells in each organ that were of the effector memory subtype. Both medians and individual data points are shown. Differences between the spleen and liver for each mouse strain were assessed using two-way ANOVA with Sidak's multiple-comparison test. For panel A, the differences were not significant; for panel B, $P < 0.0001$ for BALB/c mice and $P = 0.0027$ for C57BL/6 mice.

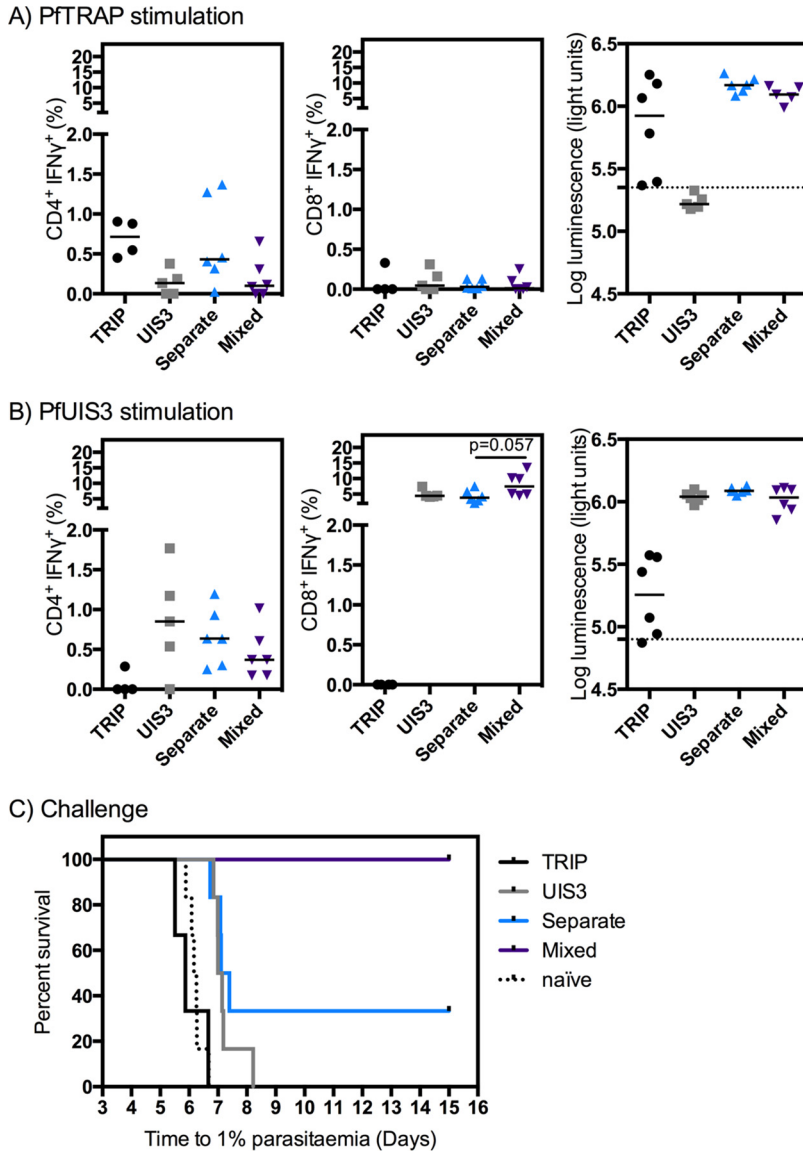


FIG 5 Coadministration of ChAd63-MVA PfUIS3 with the vaccine ChAd63-MVA TRIP is not detrimental to immunogenicity and increases efficacy in BALB/c mice. BALB/c mice ($n = 3$ to 6 per group) were vaccinated i.m. with 1×10^8 IFU of ChAd63 followed 8 weeks later by 1×10^7 PFU of MVA, with the antigens indicated on the x axis. When two vaccines were given, mice were vaccinated with a full dose of each vaccine administered in separate legs (separate), or the vaccines were mixed prior to administration in both legs (mixed). Cellular immunogenicity in the blood and IgG responses in plasma were assessed at 1 week postboost. Cells or plasma was incubated with a peptide pool or protein lysate for either PfTRAP (A) or PfUIS3 (B). Both medians and individual data points are shown. The Kruskal-Wallis test with Dunn's multiple-comparison test was used to assess statistical differences between coadministered vaccines and those given alone; no significant differences were found. (C) The same mice were then challenged at 10 days postboost with 1,000 double chimeric *P. berghei*-*P. falciparum* sporozoites that expressed both PfTRAP and PfUIS3. The log-rank (Mantel-Cox) test was used to determine the statistical difference between survival curves: for the PfTRIP versus naïve groups, the difference was not significant ($P = 0.5$); for the PfUIS3 versus naïve groups, $P = 0.0005$; for the PfUIS3 versus separate groups, the difference was not significant ($P = 0.26$); and for the PfUIS3 versus mixed groups, $P = 0.0005$.

was a higher level of variation in the TRAP-specific IgG response between mice following single TRIP vaccination than following coadministration; this likely was due to experimental variation and may account for the trend toward higher responses in the UIS3/TRAP coadministration groups. The pattern was similar for PfUIS3: slightly lower CD4⁺ IFN- γ ⁺ responses were observed following coadministration, but CD8⁺ IFN- γ ⁺ and antibody responses were higher than or equal to those with the vaccine administered alone (Fig. 5B).

To assess the protective efficacy of combination vaccination, we generated a double chimeric *P. berghei*-*P. falciparum* parasite line expressing both PfUIS3 and PfTRAP (Fig. S2 and S3). The *P. falciparum* trap and *uis3* coding sequences (CDS) were introduced as additional copies in two neutral loci in the *P. berghei* genome (*Pb230p* and *Pbs1*). Both CDS are under the control of the *Pbuis4* regulatory sequences to drive expression in sporozoites and liver-stage parasites (9). Despite the non-statistically significant immunological differences with separate and mixed PfUIS3 and PfTRIP vaccinations, these immunized mice showed improved protective efficacy against challenge with the double chimeric *P. berghei*-*P. falciparum* parasites (Fig. 5C). While PfUIS3 alone provided a delay in the time to 1% parasitemia, as expected, coadministration of PfUIS3 with PfTRIP at the same site resulted in 100% sterile protection (a significant improvement compared to the result with PfUIS3 alone [$P = 0.0005$ by the log-rank {Mantel-Cox} test] or compared to TRIP alone). There was no statistically significant improvement in efficacy when mice were immunized with both PfUIS3 and PfTRIP with delivery of the vaccines at separate sites ($P = 0.26$ compared to PfUIS3 alone) (Fig. 5C).

PfUIS3-specific memory responses are induced in human volunteers who undergo CHMI with *P. falciparum* sporozoites. We previously reported that modest *P. falciparum*-specific cellular responses are detectable following controlled human malaria infection (CHMI) with cryopreserved *P. falciparum* sporozoites (23). We did not identify one antigen that was immunodominant among the 16 tested, but each antigen did induce a modest response in at least one volunteer at one time point. As these *ex vivo* responses were of low magnitude, we aimed to determine the presence of PfUIS3-specific T cell central memory responses by using a cultured IFN- γ -specific enzyme-linked immunosorbent spot (ELISpot) assay (24). Frozen peripheral blood mononuclear cells (PBMCs) collected from 18 volunteers 35 days after CHMI with Sanaria PfSPZ challenge were stimulated for 10 days with recombinant human interleukin-2 (IL-2) and the appropriate peptide pool. Responses to PfUIS3 and four other preerythrocytic antigens (PfTRAP, PfFalstatin, PfLSA1, and PfETRAMP5) were assessed. In cases where sufficient PBMCs were available, responses were measured for all 18 volunteers, regardless of whether they were diagnosed with malaria (four volunteers did not develop blood-stage infection, as demonstrated by persistently negative quantitative PCR [qPCR] results for blood post-CHMI [23]). This was because the four volunteers who were not diagnosed with blood-stage malaria did demonstrate responses to preerythrocytic and blood-stage antigens in the *ex vivo* ELISpot assay (23; R. J. Longley and A. J. Spencer, unpublished data). Among the responses to all the antigens, only the median response to PfUIS3 fell above a positivity threshold set based upon pre-CHMI background levels, although there were no statistically significant differences in the responses to all five antigens ($P = 0.51$ by the Kruskal-Wallis test with Dunn's multiple-comparison test) (Fig. 6). If the four undiagnosed volunteers were excluded, the median response to PfUIS3 still remained higher than those to the other antigens; however, the median response to PfETRAMP5 also exceeded the positivity threshold in this case (data not shown).

DISCUSSION

Immunization with PfUIS3 administered in the viral vectors ChAd63 and MVA is unable to confer high levels of sterile protection in mice (9). However, we now demonstrate that protective immune responses can be induced in two strains of inbred mice, as shown by a consistent delay of more than 1 day in the time to 1% parasitemia after challenge with chimeric *P. berghei*-*P. falciparum* parasites expressing PfUIS3. We observed a greater level of efficacy in C57BL/6 mice than in BALB/c mice, with a 2.9-day median delay in the time to 1% parasitemia versus 1.4 days, even though C57BL/6 mice are conventionally considered more difficult to protect (25). Furthermore, after immunization with PfUIS3 in our previous study, we also observed a 1.1-day median delay in the time to 1% parasitemia in outbred mice that were challenged with the same chimeric *P. berghei*-*P. falciparum* parasites (although this was not statistically significant) (9). We show here that vaccination with PfUIS3 can also induce comparable levels of

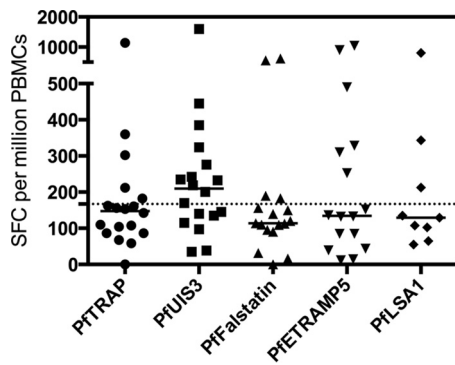


FIG 6 PfUIS3-specific memory responses were induced in human volunteers who had recently undergone CHMI with cryopreserved *P. falciparum* sporozoites. Frozen PBMCs from 18 volunteers 35 days after CHMI were stimulated for 10 days with a pool of overlapping peptides to one of five preerythrocytic antigens (PfTRAP, PfUIS3, PFI0580c, PFE1590w, and PflSA1) and recombinant human IL-2. For some volunteers, there were not enough PBMCs available to perform stimulations with all five antigens. The stimulated cells were then used in a standard IFN- γ ELISpot assay, and samples were tested in quadruplicate. Responses to antigens are expressed as numbers of spot-forming cells (SFC) per million original PBMCs. The positivity cutoff (167 SFC) was set using PBMCs from four volunteers from the day before CHMI, as described in Materials and Methods. Medians and individual data points are shown.

protection against challenge of BALB/c mice with WT *P. berghei* sporozoites, which express only PbUIS3. This is one of only a few reported cases of cross-species protection induced by malaria vaccines, in particular using subunit vaccination (21, 22, 26–30), and suggests that this vaccination regimen might be capable of providing some degree of strain-transcending immunity (at least against *P. berghei*). It was beyond the scope of the current study to determine the mechanism of cross-species protection; however, given the lack of long (8-mer) stretches of conserved residues between PfUIS3 and PbUIS3, it is possible that the protection against WT challenge is mediated by antibodies rather than cellular responses. The mechanism will be investigated in future research through the use of passive transfer experiments and mapping of cellular responses against PbUIS3.

Evidence suggests that protective immune responses after immunization with whole, attenuated parasites are dependent on (31, 32), or at least associated with (33, 34), liver-resident effector memory CD8⁺ T cells secreting IFN- γ . As the collection and analysis of liver-resident T cells are not possible in human trials, assessment of correlates of protection has largely been performed on T cells in the periphery. Studies of mice have shown that immunization with viral vectors expressing ME-TRAP induces high percentages of antigen-specific CD8⁺ T cells in the liver prior to sporozoite challenge (35, 36). Upon challenge, this enables a fast and efficient immune response to be mounted, which may be critical given the relatively short time that the parasite spends in the liver. Following PfUIS3 immunization, we observed a high percentage (10 to 20%) of PfUIS3-specific CD8⁺ T cells secreting IFN- γ in the liver for both BALB/c and C57BL/6 mice. By analysis of protective efficacy in immunized BALB/c mice that were depleted of T cells prior to challenge, we were also able to demonstrate that CD8⁺ T cells are required for protection (depletion of CD8⁺ T cells abolished the protective effect of the vaccine). In addition to this clear role of CD8⁺ T cells, we found a moderate dependence on CD4⁺ T cells: depletion of this subset significantly reduced the protective efficacy of the vaccine, but compared to control naive mice, the CD4⁺ T cell-depleted mice showed low levels of protective efficacy, as measured by a 1-day median delay in the time to 1% parasitemia in the blood. Interestingly, the conserved epitope we previously identified which induced a CD4⁺ response (amino acids 51 to 80, covering part of the region predicted to be outside the membrane) is common to both BALB/c and C57BL/6 mice, in addition to HLA-A2-transgenic mice (20). It may be that this CD4⁺ epitope contributes to the consistent level of protection seen across different mouse strains.

There is now general consensus that an effective malaria vaccine will need to contain multiple antigens, either through whole-parasite vaccination or through inclusion of multiple antigens in a subunit vaccine (37). Viral vectors offer an effective platform for this purpose (38). However, evidence suggests that combining subunit vaccines can create competition or immune interference between antigens and reduce overall immunogenicity (39, 40). The competition seen is complex, and interference seems to be antigen dependent, as other studies have combined vaccines and reported no competition (41, 42). Research has also suggested that delivery of vaccines at separate sites can reduce antigen interference (39), but again, conflicting results have been reported (40). We found no significant decline or increase in cellular or humoral immunogenicity to either antigen when PfUIS3 and PfTRIP were administered alone or in combination (whether at separate sites or mixed) in BALB/c mice. Interestingly, coadministration of the two vaccines had a synergistic effect on protective efficacy. PfTRIP alone provided no protection, and PfUIS3 alone provided a delay in patent parasitemia, as expected, but coadministration (mixed) provided 100% sterile protection. This synergistic effect was evident only when the two vaccines were mixed and administered at the same site; the efficacy was only partially improved when the vaccines were administered at separate sites. As there was no significant change in immunogenicity detected in the blood for either vaccine upon coadministration, it is difficult to know the cause of the improved outcome. However, we did observe a trend toward higher PfTRAP-specific antibody responses (mixed vaccines or separate sites) and higher PfUIS3-specific CD8⁺ responses (mixed vaccines only), which may contribute to the synergistic effect observed. A study of coadministration of a virally vectored vaccine targeting internal antigens of the influenza virus with various protein formulations of the surface antigen hemagglutinin (HA) showed higher antibody responses against HA when the two vaccines were mixed rather than administered at separate sites (43) and attributed this to local effects in draining lymph nodes (an increase in the number of germinal center B cells). It will therefore be of interest to look at immunogenicity in our model at local sites, such as the draining lymph nodes, in addition to immunogenicity in the liver, given that it is the site mediating protection.

We previously identified PfUIS3-specific epitopes in HLA-A2-transgenic mice (including the conserved CD4⁺ epitope) (20), suggesting that human populations are also capable of inducing PfUIS3-specific immunity. We have now demonstrated that human populations do indeed respond to PfUIS3: cellular memory responses were identified in human volunteers who underwent CHMI with *P. falciparum* sporozoites. The median response of volunteers against PfUIS3 was greater than those observed against the four other preerythrocytic antigens tested (albeit not statistically significantly). In past studies of nonimmune humans exposed to sporozoites (whether for CHMI or for vaccination purposes), it has been difficult to pinpoint cellular responses to a particular antigen or antigens, including in our own recent work (23, 44). This is due to a combination of factors: T cell responses induced in humans following exposure to *Plasmodium* are low and are likely directed at a wide variety of antigens (44). One way to overcome the low responses recalled *ex vivo* is to use a cultured ELISpot assay to measure memory responses (45), as we have done in this study. Apart from their different sensitivities, the *ex vivo* and cultured ELISpot assays measure different cell populations: responses measured *ex vivo* reflect effector memory cells, while those measured after an extended culture period reflect central memory cells (24). Long-term memory is a goal of all vaccination strategies, and hence the ability to induce PfUIS3-specific central memory T cells in humans is promising for the use of this antigen in a multicomponent vaccine.

In conclusion, in this study we extended our recent findings to demonstrate that vaccination with ChAd63-MVA PfUIS3 can provide a consistent level of protection across multiple strains of mice and against challenge with chimeric *P. berghei*-*P. falciparum* parasites expressing PfUIS3, as well as WT *P. berghei* (expressing only PbUIS3). This protection is dependent upon the presence of CD8⁺ T cells, and to some extent CD4⁺ T cells, for which we previously identified a conserved epitope across

multiple major histocompatibility complexes. We have also shown that humans who undergo CHMI with PfSPZ challenge develop central memory T cells against PfUIS3 and that administering PfUIS3 with PfTRIP in mice does not significantly reduce cellular immunogenicity to either antigen; in fact, combining both antigens results in 100% protection against double chimeric parasite challenge. While other antigens have been observed to provide greater levels of individual sterile protection preclinically, we have demonstrated the potential value of including PfUIS3 in a multicomponent malaria vaccine, particularly in combination with the PfTRAP antigen.

MATERIALS AND METHODS

Animals and ethics statement. Female BALB/c (H-2^d) or C57BL/6 (H-2^b) mice of at least 6 weeks of age (Harlan, United Kingdom) were used in accordance with the UK Animals (Scientific Procedures) Act 1986 and approved by the University of Oxford Animal Care and Ethical Review Committee for use under protocol PPL 30/2414 or 30/2889. Animals were group housed in individually ventilated cages under specific-pathogen-free conditions. Temperature and humidity were kept constant, and a 12-h–12-h light-dark cycle was used. Short-term anesthesia was performed using vaporized IsoFlo, and all animals were humanely sacrificed at the end of experiments by an approved schedule 1 method (cervical dislocation). All efforts were made to minimize suffering. Carrying on from our previous studies, BALB/c mice were used as the primary strain of mice for immunogenicity and efficacy; C57BL/6 mice were used in one challenge experiment to demonstrate efficacy in another inbred strain of mice and in a separate experiment to measure CD8⁺ T cell responses in the liver.

Vaccines, immunizations, and peptides. The generation of the PfUIS3 vaccine has previously been described in detail (9). Briefly, the PfUIS3 vaccine is based on the 3D7 sequence, and no modifications were made apart from optimization to mammalian codon usage bias for expression in human cells (see Fig. S1 in the supplemental material). The regions of the UIS3 protein that are immunogenic in humans are currently unknown, and hence the whole protein is still under assessment. For the PfTRAP protein, we chose to use the vaccine denoted PfTRIP rather than the clinical ME-TRAP construct. This was for the following two reasons: first, the ME string contains the strong *P. berghei* Pb9 H-2^d-restricted epitope from CSP (46), and hence this cannot be used to measure immunogenicity and efficacy directed at the TRAP protein in BALB/c mice; and second, the PfTRIP construct uses the 3D7 sequence (PlasmoDB accession no. PF3D7_1335900) (ME-TRAP uses the T9/96 strain sequence [47]), as does the PfUIS3 construct and the challenge parasites. The PfTRIP vaccine was codon optimized for expression in human cells and had 15 amino acids deleted, as does ME-TRAP (five repeats of PNP), as well as deletion of the predicted transmembrane helix and cytoplasmic domains (Fig. S1). The first 300 amino acids of the TRAP protein are known to be more immunogenic in humans than the transmembrane helix and cytoplasmic domains (48). Mice were immunized intramuscularly (i.m.) with 50 μ l vaccine (in endotoxin-free Dulbecco's phosphate-buffered saline [D-PBS]) into the musculus tibialis, with doses stated in the relevant figure legends. When two vaccines were coadministered, full doses were administered individually into separate legs or mixed and administered into both legs ($2 \times 50 \mu$ l). All vaccines were administered using a ChAd63 prime followed 8 weeks later by an MVA boost. Overlapping peptide pools (synthetic crude 20-mers overlapping by 10 amino acids) were used to stimulate cells in the various immunoassays described below. The PfUIS3 pool contained a mix of 22 peptides (20), and the PfTRAP pool contained 50 peptides (Table S2). Each individual peptide was originally dissolved to a concentration of either 50 or 100 mg/ml prior to pooling of the peptides for use in *in vitro* restimulation at a final peptide concentration of 5 μ g/ml. The dimethyl sulfoxide (DMSO) concentration was always less than 1% in the final peptide pool. Peptides were purchased from Neo Group Inc. or Thermo Fisher Scientific.

ICS assay. An intracellular cytokine staining (ICS) assay was used to detect antigen-specific cellular responses in the blood, spleen, and liver and was performed as previously described (9). Briefly, after lysis of red blood cells, single-cell suspensions were stimulated for 6 h with a final concentration of 5 μ g/ml of the appropriate peptide pool (PfUIS3 or PfTRAP), 1 μ g/ml GolgiPlug (BD Biosciences, United Kingdom), and anti-mouse CD107a–phycoerythrin (PE). Unstimulated wells without peptide were used as background controls. For standard ICS, cells were surface stained with anti-mouse CD16/32, CD4-eFluor 450, and CD8 α –peridinin chlorophyll protein (PerCP)–Cy5.5 antibodies, and intracellular cytokines were stained with tumor necrosis factor alpha (TNF- α)–fluorescein isothiocyanate (FITC), IL-2–PE–Cy7, and IFN- γ –allophycocyanin (APC) antibodies diluted in Perm/Wash reagent (BD Biosciences). The gating strategy and representative plots are shown in Fig. S2 in the supplemental material. To stain for memory cell markers, the first layer contained anti-mouse CD16/32, CD8 α –PerCP–Cy5.5, CD4-eFluor 650, CD62L–PE–Cy7, and CD127–APC–eFluor 780 antibodies and Live/Dead Aqua reagent. The second layer contained anti-mouse TNF- α –FITC and IFN- γ –eFluor 450 antibodies diluted in Perm/Wash reagent. The gating strategy used for memory cell markers was published previously (9). Data were acquired using an LSRII flow cytometer (BD Biosciences) and analyzed using FlowJo (Tree Star Inc.). All antibodies were purchased from BD Biosciences, eBioscience, or Invitrogen, United Kingdom.

Antibody measurements. IgG antibody responses were measured using the luminescence immunoprecipitation system (LIPS) as previously described (9).

Parasites and efficacy studies. Sporozoites were obtained by dissection and homogenization of salivary glands from *Anopheles stephensi* mosquitoes at 21 days postinfection. Blood stocks of green fluorescent protein (GFP)-expressing *P. berghei* parasites (49) were kindly provided by Robert Sinden (Imperial College, London, and Oxford University). The generation of chimeric *P. berghei*-*P. falciparum*

parasites expressing PfUIS3 has been described previously (9). In addition, we generated double chimeric *P. berghei*-*P. falciparum* parasites that express both PfUIS3 and PfTRAP under the control of the *Pbuis4* promoter (see below). One thousand sporozoites were injected intravenously (i.v.) into the tail vein 8 to 10 days following the final vaccination, as detailed in the relevant figure legends. Mice were monitored from 4 days postinfection by use of Giemsa-stained thin-film blood smears. The experimental endpoint was 14 or 15 days parasite free (sterilely protected) or when blood-stage parasites had been confirmed on three consecutive days. The time to 1% parasitemia was calculated using linear regression; when sterile protection is not achieved, this value is adept at providing a sensitive measure of liver-stage protection, as it reflects the number of parasites erupting from the liver (50, 51). In some experiments, subsets of T cells were depleted using the monoclonal antibody (MAb) anti-CD4 clone GK1.5 or anti-CD8 clone 2.43 (both are rat IgG2a antibodies purified by protein G affinity chromatography from hybridoma culture supernatants). An IgG control (Sigma-Aldrich) was purified using the same method. One hundred micrograms of antibody injected 2 days before and on the day of challenge successfully depleted 100% of either cell population (9), and this depletion was maintained until at least 4 days postchallenge. The degree of *in vivo* CD4⁺ or CD8⁺ T cell depletion was assessed by flow cytometry using 1/100 anti-CD4-FITC clone RM4-5, 1/200 anti-CD8-PerCP-Cy5.5 clone 53-6.7, and 1/50 anti-CD3 ϵ -APC.

Generation of *P. berghei*-*P. falciparum* double chimeric parasites expressing both PfUIS3 and PfTRAP. To generate the double additional gene (DAG) chimeric parasite PbANKA-PfTRAP+PfUIS3@Pbuis4 DAG (line 2395c1), we used the previously generated single additional gene (SAG) PbANKA-PfTRAP_{Pbuis4} line (line 2281c1) as the background parent line (9). This line expresses the PfTRAP (accession no. PF3D7_1335900) coding sequence (CDS) under the control of the *Pbuis4* regulatory sequences and a fusion protein of GFP and firefly luciferase (LUC-IAV) under the control of the constitutive *Pbeef1a* promoter. This transgenic parasite is selectable marker (SM) free. Both the *trap* and *Luc-gfp* expression cassettes are integrated into the neutral 230p locus in chromosome 3. In this line, the PfUIS3 CDS (PfUIS3; accession no. PF3D7_1302200) was stably integrated into a neutral *s1* gene locus (*Pbs1*; accession no. PBANKA_120680) (52, 53; A. M. Salman and C. J. Janse, unpublished data) through double-crossover recombination using a 2-step "gene insertion/marker out" (GIMO) transfection protocol (52, 54) (Fig. S3). In the first step, we deleted the *Pbs1* CDS and replaced it with a positive-negative selectable marker to create a *Pbs1* deletion GIMO line (PbANKA-PfTRAP+Pb Δ s1 GIMO; line 2353c12). In order to do this, we generated the pL1928 construct, based on the standard GIMO DNA construct pL0034 (54). This construct contains a positive-negative (*hdhfr::yfcu*) SM cassette and was used to insert both the *Pbs1* 5' and 3' gene targeting regions (TRs) (52). The linear pL1928 DNA construct was introduced into PbANKA-PfTRAP_{Pbuis4} parasites (line 2281c1) (9) by using standard methods of transfection (55). Transfected parasites were selected in mice through addition of pyrimethamine in the drinking water (55). Transfected parasites were cloned by limiting dilution (56), resulting in the PbANKA-PfTRAP+Pb Δ s1 GIMO line (line 2353c12). Correct deletion of the *Pbs1* CDS and its replacement with the SM cassette were confirmed by diagnostic PCR analysis of genomic DNA (gDNA) and Southern analysis of pulsed-field gel electrophoresis-separated chromosomes as described previously (9) (Fig. S4). Primers used for PCR genotyping are listed in Table S2.

In the second step, we replaced the positive-negative SM in the genome of the PbANKA-PfTRAP+Pb Δ s1 line (line 2353c12) with the PfUIS3 CDS by GIMO transfection to create a *P. berghei* DAG chimeric line expressing PfTRAP and PfUIS3. This was achieved by modifying the construct used in the first step (pL1928); specifically, the *hdhfr::yfcu* SM cassette was removed and replaced with the PfUIS3 CDS expression cassette, generating plasmid pL2043. The expression cassette in pL2043 contained the PfUIS3 CDS flanked by the 5' and 3' regulatory sequences of *Pbuis4*, which were amplified from *P. berghei* ANKA WT genomic DNA. The PfUIS3 CDS was amplified from genomic DNA of the NF54 strain of *P. falciparum*. The pL2043 construct was sequenced to ensure that there were no mutations in the PfUIS3 CDS. The construct was linearized using the HindIII restriction enzyme sites outside the 5' and 3' *Pbs1* TRs before transfection. The construct was used to transfect parasites of the PbANKA-PfTRAP+Pb Δ s1 GIMO line (line 2353c12) by using standard methods of GIMO transfection (54). Transfected parasites were selected in mice by applying negative selection by providing 5-fluorocytosine (5FC) in the drinking water (57). This resulted in selection of chimeric parasites in which the *hdhfr::yfcu* SM in the *Pbs1* locus of the PbANKA-PfTRAP+Pb Δ s1 GIMO line was replaced by the PfUIS3 CDS expression cassette. Selected chimeric parasites were cloned by limiting dilution (56). Correct integration of the constructs into the genome of chimeric parasites was analyzed by diagnostic PCR analysis of gDNA and Southern analysis of pulsed-field gel electrophoresis-separated chromosomes. Primers used for PCR genotyping are listed in Table S1. This method created DAG chimeric *P. berghei*-*P. falciparum* parasites that express full-length PfTRAP and PfUIS3 under the control of the *Pbuis4* regulatory sequences and are SM free.

Phenotyping of chimeric parasites. Multiplication of blood stages in mice was determined during the cloning period as described previously (58). Feeding of *A. stephensi* mosquitoes for determination of oocyst and sporozoite numbers was performed as described previously (9). Infection dynamics of the double chimeric parasites in mice following sporozoite injection were equivalent to those of WT *P. berghei* parasites (Fig. S5).

CHMI with PfSPZ challenge and cultured IFN- γ ELISpot assay. Cryopreserved human peripheral blood mononuclear cells (PBMCs) were obtained from a previously published clinical trial of the Sanaria PfSPZ challenge in malaria-naive UK adults (trial no. NCT01465048 at ClinicalTrials.gov) (23). In brief, 18 healthy volunteers received aseptic, purified, cryopreserved, live, infectious *P. falciparum* sporozoites (PfSPZ challenge [59]) (2,500 or 25,000) injected either i.m. or intradermally. A total of 14/18 volunteers had parasites identified in the blood by quantitative PCR performed postinjection. Cryopreserved PBMCs from 35 days postinjection were used to assess T cell memory responses in a cultured IFN- γ ELISpot assay

(24). A total of 5×10^6 thawed cells were added to the middle wells of a 12-well plate, along with $4 \mu\text{g/ml}$ of the appropriate peptide pool, and incubated at 37°C with $5\% \text{CO}_2$. On days 3 and 7, half the medium volume was replaced with recombinant human IL-2 (final concentration, 25 IU/ml). On day 9, cells were washed to remove all traces of IL-2 and left to rest overnight. On day 10, the IFN- γ ELISpot assay was performed as previously described (23). Briefly, cells were incubated for 18 to 20 h with $20 \mu\text{g/ml}$ of the appropriate overlapping peptide pool in ELISpot plates (Mabtech, Sweden) coated with anti-IFN- γ . After addition of the detecting antibody and substrate, spots were enumerated using an ELISpot plate counter (AID, Germany). All wells were set up in quadruplicate. Due to the higher background and variability in a cultured versus *ex vivo* ELISpot assay, a stringent positivity cutoff was set by performing the assay using cryopreserved PBMCs taken from volunteers the day prior to controlled human malaria infection (CHMI) with PfSPZ challenge. This was performed for four volunteers with two different antigenic stimulations, and the positivity cutoff was set as the overall mean plus 2 standard deviations.

Statistical analysis. The statistical software Prism, version 6 (GraphPad), was used for all analyses. Survival in challenge experiments is presented using Kaplan-Meier curves, and significance was tested using the log-rank (Mantel-Cox) test. The significance threshold was 0.05. Nonparametric data are shown as medians with individual data points plotted unless otherwise indicated.

SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at <https://doi.org/10.1128/IAI.00641-16>.

TEXT S1, PDF file, 1.1 MB.

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A.V.S.H. is a named investigator on novel adenovirus patent applications U.S. 12/595 574 and UK PCT/GB2008/01262, covering malaria vectored vaccines and immunization regimens. R.J.L., A.J.S., and A.V.S.H. are named investigators on patent PCT/GB2014/053077, identifying novel malaria vaccine antigens.

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REFERENCES

1. Plowe CV, Alonso P, Hoffman SL. 2009. The potential role of vaccines in the elimination of falciparum malaria and the eventual eradication of malaria. *J Infect Dis* 200:1646–1649. <https://doi.org/10.1086/646613>.
2. Cohen J, Nussenzweig V, Nussenzweig R, Vekemans J, Leach A. 2010. From the circumsporozoite protein to the RTS, S/AS candidate vaccine. *Hum Vaccin* 6:90–96. <https://doi.org/10.4161/hv.6.1.9677>.
3. Anonymous. 2015. Efficacy and safety of RTS,S/AS01 malaria vaccine with or without a booster dose in infants and children in Africa: final results of a phase 3, individually randomised, controlled trial. *Lancet* 386:31–45. [https://doi.org/10.1016/S0140-6736\(15\)60721-8](https://doi.org/10.1016/S0140-6736(15)60721-8).
4. Reyes-Sandoval A, Rollier CS, Milicic A, Bauza K, Cottingham MG, Tang CK, Dicks MD, Wang D, Longley RJ, Wyllie DH, Hill AV. 2012. Mixed vector immunization with recombinant adenovirus and MVA can improve vac-

- line efficacy while decreasing antivector immunity. *Mol Ther* 20: 1633–1647. <https://doi.org/10.1038/mt.2012.25>.
5. Ewer KJ, O'Hara GA, Duncan CJ, Collins KA, Sheehy SH, Reyes-Sandoval A, Goodman AL, Edwards NJ, Elias SC, Halstead FD, Longley RJ, Rowland R, Poulton ID, Draper SJ, Blagborough AM, Berrie E, Moyle S, Williams N, Siani L, Folgori A, Colloca S, Sinden RE, Lawrie AM, Cortese R, Gilbert SC, Nicosia A, Hill AV. 2013. Protective CD8+ T-cell immunity to human malaria induced by chimpanzee adenovirus-MVA immunisation. *Nat Commun* 4:2836. <https://doi.org/10.1038/ncomms3836>.
 6. Trimnell A, Takagi A, Gupta M, Richie TL, Kappe SH, Wang R. 2009. Genetically attenuated parasite vaccines induce contact-dependent CD8+ T cell killing of *Plasmodium yoelii* liver stage-infected hepatocytes. *J Immunol* 183:5870–5878. <https://doi.org/10.4049/jimmunol.0900302>.
 7. O'Hara GA, Duncan CJ, Ewer KJ, Collins KA, Elias SC, Halstead FD, Goodman AL, Edwards NJ, Reyes-Sandoval A, Bird P, Rowland R, Sheehy SH, Poulton ID, Hutchings C, Todryk S, Andrews L, Folgori A, Berrie E, Moyle S, Nicosia A, Colloca S, Cortese R, Siani L, Lawrie AM, Gilbert SC, Hill AV. 2012. Clinical assessment of a recombinant simian adenovirus ChAd63: a potent new vaccine vector. *J Infect Dis* 205:772–781. <https://doi.org/10.1093/infdis/jir850>.
 8. Ogwang C, Kimani D, Edwards NJ, Roberts R, Mwacharo J, Bowyer G, Bliss C, Hodgson SH, Njuguna P, Viebig NK, Nicosia A, Gitau E, Douglas S, Illingworth J, Marsh K, Lawrie A, Imoukhuede EB, Ewer K, Urban BC, Hill AVS, Bejon P, MVVC Group. 2015. Prime-boost vaccination with chimpanzee adenovirus and modified vaccinia Ankara encoding TRAP provides partial protection against *Plasmodium falciparum* infection in Kenyan adults. *Sci Transl Med* 7:286re285. <https://doi.org/10.1126/scitranslmed.aaa2373>.
 9. Longley RJ, Salman AM, Cottingham MG, Ewer K, Janse CJ, Khan SM, Spencer AJ, Hill AV. 2015. Comparative assessment of vaccine vectors encoding ten malaria antigens identifies two protective liver-stage candidates. *Sci Rep* 5:11820. <https://doi.org/10.1038/srep11820>.
 10. Spielmann T, Fergusen DJ, Beck HP. 2003. etramps, a new *Plasmodium falciparum* gene family coding for developmentally regulated and highly charged membrane proteins located at the parasite-host cell interface. *Mol Biol Cell* 14:1529–1544. <https://doi.org/10.1091/mbc.E02-04-0240>.
 11. Matuschewski K, Ross J, Brown SM, Kaiser K, Nussenzweig V, Kappe SH. 2002. Infectivity-associated changes in the transcriptional repertoire of the malaria parasite sporozoite stage. *J Biol Chem* 277:41948–41953. <https://doi.org/10.1074/jbc.M207315200>.
 12. Kaiser K, Matuschewski K, Camargo N, Ross J, Kappe SH. 2004. Differential transcriptome profiling identifies *Plasmodium* genes encoding pre-erythrocytic stage-specific proteins. *Mol Microbiol* 51:1221–1232. <https://doi.org/10.1046/j.1365-2958.2003.03909.x>.
 13. Limbach K, Aguiar J, Gowda K, Patterson N, Abot E, Sedegah M, Sacchi J, Richie T. 2011. Identification of two new protective pre-erythrocytic malaria vaccine antigen candidates. *Malar J* 10:65. <https://doi.org/10.1186/1475-2875-10-65>.
 14. Le Roch KG, Zhou Y, Blair PL, Grainger M, Moch JK, Haynes JD, De La Vega P, Holder AA, Batalov S, Carucci DJ, Winzeler EA. 2003. Discovery of gene function by expression profiling of the malaria parasite life cycle. *Science* 301:1503–1508. <https://doi.org/10.1126/science.1087025>.
 15. Mueller AK, Labaied M, Kappe SH, Matuschewski K. 2005. Genetically modified *Plasmodium* parasites as a protective experimental malaria vaccine. *Nature* 433:164–167. <https://doi.org/10.1038/nature03188>.
 16. Tarun AS, Dumpit RF, Camargo N, Labaied M, Liu P, Takagi A, Wang R, Kappe SH. 2007. Protracted sterile protection with *Plasmodium yoelii* pre-erythrocytic genetically attenuated parasite malaria vaccines is independent of significant liver-stage persistence and is mediated by CD8+ T cells. *J Infect Dis* 196:608–616. <https://doi.org/10.1086/519742>.
 17. Mikolajczak SA, Jacobs-Lorena V, MacKellar DC, Camargo N, Kappe SH. 2007. L-FABP is a critical host factor for successful malaria liver stage development. *Int J Parasitol* 37:483–489. <https://doi.org/10.1016/j.ijpara.2007.01.002>.
 18. Sharma A, Yogavel M, Akhouri RR, Gill J, Sharma A. 2008. Crystal structure of soluble domain of malaria sporozoite protein UIS3 in complex with lipid. *J Biol Chem* 283:24077–24088. <https://doi.org/10.1074/jbc.M801946200>.
 19. Storch J, Thumser AE. 2010. Tissue-specific functions in the fatty acid-binding protein family. *J Biol Chem* 285:32679–32683. <https://doi.org/10.1074/jbc.R110.135210>.
 20. Longley RJ, Halbroth BR, Ewer KJ, Hill AV, Spencer AJ. 2015. Identification of immunodominant responses to the *Plasmodium falciparum* antigens PfUIS3, PflSA1 and PflSA2 in multiple strains of mice. *PLoS One* 10:e0144515. <https://doi.org/10.1371/journal.pone.0144515>.
 21. Sauzet JP, Perlaza BL, Brahimi K, Daubersies P, Druilhe P. 2001. DNA immunization by *Plasmodium falciparum* liver-stage antigen 3 induces protection against *Plasmodium yoelii* sporozoite challenge. *Infect Immun* 69:1202–1206. <https://doi.org/10.1128/IAI.69.2.1202-1206.2001>.
 22. Yadava A, Nurmukhambetova S, Pichugin AV, Lumsden JM. 2012. Cross-species immunity following immunization with a circumsporozoite protein-based vaccine for malaria. *J Infect Dis* 205:1456–1463. <https://doi.org/10.1093/infdis/jis220>.
 23. Sheehy SH, Spencer AJ, Douglas AD, Sim BK, Longley RJ, Edwards NJ, Poulton ID, Kimani D, Williams AR, Anagnostou NA, Roberts R, Kerridge S, Voysey M, James ER, Billingsley PF, Gunasekera A, Lawrie AM, Hoffman SL, Hill AV. 2013. Optimising controlled human malaria infection studies using cryopreserved parasites administered by needle and syringe. *PLoS One* 8:e65960. <https://doi.org/10.1371/journal.pone.0065960>.
 24. Todryk SM, Pathan AA, Keating S, Porter DW, Berthoud T, Thompson F, Klenerman P, Hill AV. 2009. The relationship between human effector and memory T cells measured by ex vivo and cultured ELISPOT following recent and distal priming. *Immunology* 128:83–91. <https://doi.org/10.1111/j.1365-2567.2009.03073.x>.
 25. Matuschewski K. 2013. Murine infection models for vaccine development: the malaria example. *Hum Vaccin Immunother* 9:450–456. <https://doi.org/10.4161/hv.23218>.
 26. Butler NS, Schmidt NW, Vaughan AM, Aly AS, Kappe SH, Harty JT. 2011. Superior antimalarial immunity after vaccination with late liver stage-arresting genetically attenuated parasites. *Cell Host Microbe* 9:451–462. <https://doi.org/10.1016/j.chom.2011.05.008>.
 27. Douradinha B, van Dijk MR, Ataide R, van Gemert GJ, Thompson J, Franetich JF, Mazier D, Luty AJ, Sauerwein R, Janse CJ, Waters AP, Mota MM. 2007. Genetically attenuated P36p-deficient *Plasmodium berghei* sporozoites confer long-lasting and partial cross-species protection. *Int J Parasitol* 37:1511–1519. <https://doi.org/10.1016/j.ijpara.2007.05.005>.
 28. Mauduit M, Gruner AC, Tewari R, Depinay N, Kayibanda M, Chavatte JM, Franetich JF, Crisanti A, Mazier D, Snounou G, Renia L. 2009. A role for immune responses against non-CS components in the cross-species protection induced by immunization with irradiated malaria sporozoites. *PLoS One* 4:e7717. <https://doi.org/10.1371/journal.pone.0007717>.
 29. Good MF, Reiman JM, Rodriguez IB, Ito K, Yanow SK, El-Deeb IM, Batzloff MR, Stanicic DI, Engwerda C, Spithill T, Hoffman SL, Lee M, McPhun V. 2013. Cross-species malaria immunity induced by chemically attenuated parasites. *J Clin Invest* 2013:66634. <https://doi.org/10.1172/jci66634>.
 30. Bergmann-Leitner ES, Mease RM, De La Vega P, Savranskaya T, Polhemus M, Ockenhouse C, Angov E. 2010. Immunization with pre-erythrocytic antigen CelTOS from *Plasmodium falciparum* elicits cross-species protection against heterologous challenge with *Plasmodium berghei*. *PLoS One* 5:e12294. <https://doi.org/10.1371/journal.pone.0012294>.
 31. Guebre-Xabier M, Schwenk R, Krzych U. 1999. Memory phenotype CD8(+) T cells persist in livers of mice protected against malaria by immunization with attenuated *Plasmodium berghei* sporozoites. *Eur J Immunol* 29:3978–3986.
 32. Morrot A, Hafalla JC, Cockburn IA, Carvalho LH, Zavala F. 2005. IL-4 receptor expression on CD8+ T cells is required for the development of protective memory responses against liver stages of malaria parasites. *J Exp Med* 202:551–560. <https://doi.org/10.1084/jem.20042463>.
 33. Berenzon D, Schwenk RJ, Letellier L, Guebre-Xabier M, Williams J, Krzych U. 2003. Protracted protection to *Plasmodium berghei* malaria is linked to functionally and phenotypically heterogeneous liver memory CD8+ T cells. *J Immunol* 171:2024–2034. <https://doi.org/10.4049/jimmunol.171.4.2024>.
 34. Nganou-Makamdop K, van Gemert GJ, Arens T, Hermsen CC, Sauerwein RW. 2012. Long term protection after immunization with *P. berghei* sporozoites correlates with sustained IFN γ responses of hepatic CD8+ memory T cells. *PLoS One* 7:e36508. <https://doi.org/10.1371/journal.pone.0036508>.
 35. Reyes-Sandoval A, Wyllie DH, Bauza K, Milicic A, Forbes EK, Rollier CS, Hill AV. 2011. CD8+ T effector memory cells protect against liver-stage malaria. *J Immunol* 187:1347–1357. <https://doi.org/10.4049/jimmunol.1100302>.
 36. Reyes-Sandoval A, Berthoud T, Alder N, Siani L, Gilbert SC, Nicosia A, Colloca S, Cortese R, Hill AV. 2010. Prime-boost immunization with adenoviral and modified vaccinia virus Ankara vectors enhances the durability and polyfunctionality of protective malaria CD8+ T-cell

- responses. *Infect Immun* 78:145–153. <https://doi.org/10.1128/IAI.00740-09>.
37. Good MF, Doolan DL. 2010. Malaria vaccine design: immunological considerations. *Immunity* 33:555–566. <https://doi.org/10.1016/j.immuni.2010.10.005>.
 38. Tine JA, Lanar DE, Smith DM, Welde BT, Schultheiss P, Ware LA, Kauffman EB, Wirtz RA, De Taisne C, Hui GS, Chang SP, Church P, Hollingdale MR, Kaslow DC, Hoffman S, Guito KP, Ballou WR, Sadoff JC, Paoletti E. 1996. NYVAC-Pf7: a poxvirus-vectored, multiantigen, multistage vaccine candidate for *Plasmodium falciparum* malaria. *Infect Immun* 64:3833–3844.
 39. Forbes EK, Biswas S, Collins KA, Gilbert SC, Hill AV, Draper SJ. 2011. Combining liver- and blood-stage malaria viral-vectored vaccines: investigating mechanisms of CD8+ T cell interference. *J Immunol* 187:3738–3750. <https://doi.org/10.4049/jimmunol.1003783>.
 40. Pichyangkul S, Tongtawe P, Kum-Arb U, Yongvanitchit K, Gettayacamin M, Hollingdale MR, Limsalakpetch A, Stewart VA, Lanar DE, Dutta S, Angov E, Ware LA, Bergmann-Leitner ES, House B, Voss G, Dubois MC, Cohen JD, Fukuda MM, Heppner DG, Miller RS. 2009. Evaluation of the safety and immunogenicity of *Plasmodium falciparum* apical membrane antigen 1, merozoite surface protein 1 or RTS,S vaccines with adjuvant system AS02A administered alone or concurrently in rhesus monkeys. *Vaccine* 28:452–462. <https://doi.org/10.1016/j.vaccine.2009.10.022>.
 41. Grifantini R, Finco O, Bartolini E, Draghi M, Del Giudice G, Kocken C, Thomas A, Abrignani S, Grandi G. 1998. Multi-plasmid DNA vaccination avoids antigenic competition and enhances immunogenicity of a poorly immunogenic plasmid. *Eur J Immunol* 28:1225–1232. [https://doi.org/10.1002/\(SICI\)1521-4141\(199804\)28:04<1225::AID-IMMU1225>3.0.CO;2-6](https://doi.org/10.1002/(SICI)1521-4141(199804)28:04<1225::AID-IMMU1225>3.0.CO;2-6).
 42. Jiang G, Charoenvit Y, Moreno A, Baraceros MF, Banania G, Richie N, Abot S, Ganeshan H, Fallarme V, Patterson NB, Geall A, Weiss WR, Strobert E, Caro-Aquilar I, Lanar DE, Saul A, Martin LB, Gowda K, Morrisette CR, Kaslow DC, Carucci DJ, Galinski MR, Doolan DL. 2007. Induction of multi-antigen multi-stage immune responses against *Plasmodium falciparum* in rhesus monkeys, in the absence of antigen interference, with heterologous DNA prime/poxvirus boost immunization. *Malar J* 6:135. <https://doi.org/10.1186/1475-2875-6-135>.
 43. Mullarkey CE, Boyd A, van Laarhoven A, Lefevre EA, Veronica Carr B, Baratelli M, Molesti E, Temperton NJ, Butter C, Charleston B, Lambe T, Gilbert SC. 2013. Improved adjuvanting of seasonal influenza vaccines: preclinical studies of MVA-NP+M1 coadministration with inactivated influenza vaccine. *Eur J Immunol* 43:1940–1952. <https://doi.org/10.1002/eji.201242922>.
 44. Epstein JE, Tewari K, Lyke KE, Sim BK, Billingsley PF, Laurens MB, Gunasekera A, Chakravarty S, James ER, Sedegah M, Richman A, Velmurugan S, Reyes S, Li M, Tucker K, Ahumada A, Ruben AJ, Li T, Stafford R, Eappen AG, Tamminga C, Bennett JW, Ockenhouse CF, Murphy JR, Komisar J, Thomas N, Loyevsky M, Birkett A, Plowe CV, Loucq C, Edelman R, Richie TL, Seder RA, Hoffman SL. 2011. Live attenuated malaria vaccine designed to protect through hepatic CD8(+) T cell immunity. *Science* 334:475–480. <https://doi.org/10.1126/science.1211548>.
 45. Aguiar JC, Bolton J, Wanga J, Sacci JB, Iriko H, Mazeika JK, Han ET, Limbach K, Patterson NB, Sedegah M, Cruz AM, Tsuboi T, Hoffman SL, Carucci D, Hollingdale MR, Villasante ED, Richie TL. 2015. Discovery of novel *Plasmodium falciparum* pre-erythrocytic antigens for vaccine development. *PLoS One* 10:e0136109. <https://doi.org/10.1371/journal.pone.0136109>.
 46. Romero P, Maryanski JL, Corradin G, Nussenzweig RS, Nussenzweig V, Zavala F. 1989. Cloned cytotoxic T cells recognize an epitope in the circumsporozoite protein and protect against malaria. *Nature* 341:323–326. <https://doi.org/10.1038/341323a0>.
 47. McConkey SJ, Reece WH, Moorthy VS, Webster D, Dunachie S, Butcher G, Vuola JM, Blanchard TJ, Gothard P, Watkins K, Hannan CM, Everaere S, Brown K, Kester KE, Cummings J, Williams J, Heppner DG, Pathan A, Flanagan K, Arulanantham N, Roberts MT, Roy M, Smith GL, Schneider J, Peto T, Sinden RE, Gilbert SC, Hill AV. 2003. Enhanced T-cell immunogenicity of plasmid DNA vaccines boosted by recombinant modified vaccinia virus Ankara in humans. *Nat Med* 9:729–735. <https://doi.org/10.1038/nm881>.
 48. Hodgson SH, Ewer KJ, Bliss CM, Edwards NJ, Rampling T, Anagnostou NA, de Barra E, Havelock T, Bowyer G, Poulton ID, de Cassan S, Longley R, Illingworth JJ, Douglas AD, Mange PB, Collins KA, Roberts R, Gerry S, Berrie E, Moyle S, Colloca S, Cortese R, Sinden RE, Gilbert SC, Bejon P, Lawrie AM, Nicosia A, Faust SN, Hill AV. 2015. Evaluation of the efficacy of ChAd63-MVA vectored vaccines expressing circumsporozoite protein and ME-TRAP against controlled human malaria infection in malaria-naïve individuals. *J Infect Dis* 211:1076–1086. <https://doi.org/10.1093/infdis/jiu579>.
 49. Franke-Fayard B, Trueman H, Ramesar J, Mendoza J, van der Keur M, van der Linden R, Sinden RE, Waters AP, Janse CJ. 2004. A *Plasmodium berghei* reference line that constitutively expresses GFP at a high level throughout the complete life cycle. *Mol Biochem Parasitol* 137:23–33. <https://doi.org/10.1016/j.molbiopara.2004.04.007>.
 50. Hafalla JC, Bauza K, Friesen J, Gonzalez-Aseguinolaza G, Hill AV, Matuschewski K. 2013. Identification of targets of CD8(+) T cell responses to malaria liver stages by genome-wide epitope profiling. *PLoS Pathog* 9:e1003303. <https://doi.org/10.1371/journal.ppat.1003303>.
 51. Bejon P, Andrews L, Andersen RF, Dunachie S, Webster D, Walther M, Gilbert SC, Peto T, Hill AV. 2005. Calculation of liver-to-blood inocula, parasite growth rates, and preerythrocytic vaccine efficacy, from serial quantitative polymerase chain reaction studies of volunteers challenged with malaria sporozoites. *J Infect Dis* 191:619–626. <https://doi.org/10.1086/427243>.
 52. Salman AM, Mogollon CM, Lin JW, van Pul FJ, Janse CJ, Khan SM. 2015. Generation of transgenic rodent malaria parasites expressing human malaria parasite proteins. *Methods Mol Biol* 1325:257–286. https://doi.org/10.1007/978-1-4939-2815-6_21.
 53. Jacobs-Lorena VY, Mikolajczak SA, Labaied M, Vaughan AM, Kappe SH. 2010. A dispensable *Plasmodium* locus for stable transgene expression. *Mol Biochem Parasitol* 171:40–44. <https://doi.org/10.1016/j.molbiopara.2009.12.009>.
 54. Lin JW, Annoura T, Sajid M, Chevalley-Maurel S, Ramesar J, Klop O, Franke-Fayard BM, Janse CJ, Khan SM. 2011. A novel 'gene insertion/marker out' (GIMO) method for transgene expression and gene complementation in rodent malaria parasites. *PLoS One* 6:e29289. <https://doi.org/10.1371/journal.pone.0029289>.
 55. Janse CJ, Franke-Fayard B, Mair GR, Ramesar J, Thiel C, Engelmann S, Matuschewski K, van Gemert GJ, Sauerwein RW, Waters AP. 2006. High efficiency transfection of *Plasmodium berghei* facilitates novel selection procedures. *Mol Biochem Parasitol* 145:60–70. <https://doi.org/10.1016/j.molbiopara.2005.09.007>.
 56. Menard R, Janse C. 1997. Gene targeting in malaria parasites. *Methods* 13:148–157. <https://doi.org/10.1006/meth.1997.0507>.
 57. Orr RY, Philip N, Waters AP. 2012. Improved negative selection protocol for *Plasmodium berghei* in the rodent malarial model. *Malar J* 11:103. <https://doi.org/10.1186/1475-2875-11-103>.
 58. Janse CJ, Ramesar J, Waters AP. 2006. High-efficiency transfection and drug selection of genetically transformed blood stages of the rodent malaria parasite *Plasmodium berghei*. *Nat Protoc* 1:346–356. <https://doi.org/10.1038/nprot.2006.53>.
 59. Epstein JE. 2013. Taking a bite out of malaria: controlled human malaria infection by needle and syringe. *Am J Trop Med Hyg* 88:3–4. <https://doi.org/10.4269/ajtmh.2013.12-0715>.